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Levi Jamison⁵, Dan Bean⁶, Ruth
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Hohenlohe¹



@Amanda_Stahlke

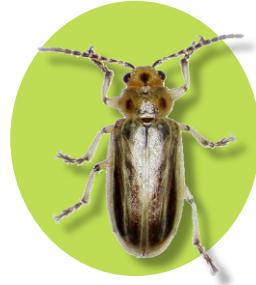
1. Institute for Bioinformatics and Evolutionary Studies, University of Idaho
2. Colorado Mesa University, Grand Junction, CO
3. Colorado State University, Fort Collins, CO
4. U.S. Department of Agriculture – Agricultural Research Service
5. EcoPlateau Research,
6. Colorado Department of Agriculture
7. Colorado Plateau Research Station, Northern Arizona University

Historically isolated tamarisk beetle species meet again on the Rio Grande & Pecos River

2019 Riparian Restoration Conference

February 6

Six source populations of *Diorhabda* spp. have been introduced since 2001



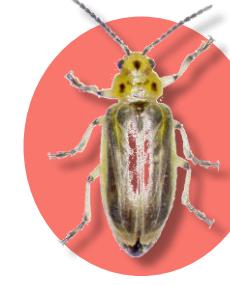
D.

sulcata



D.

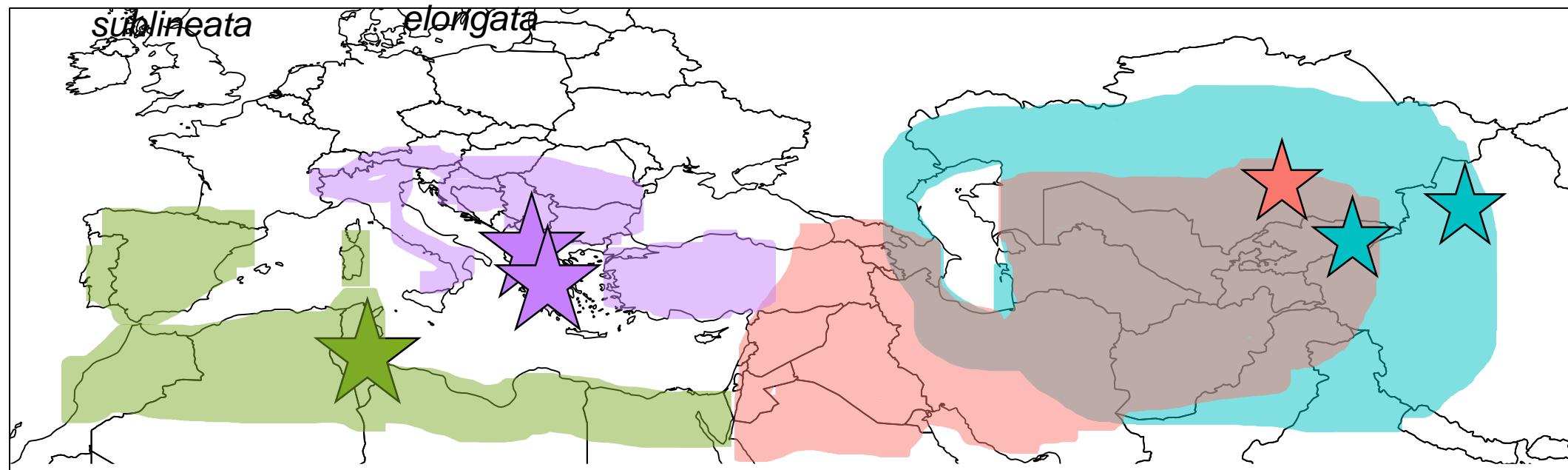
elongata



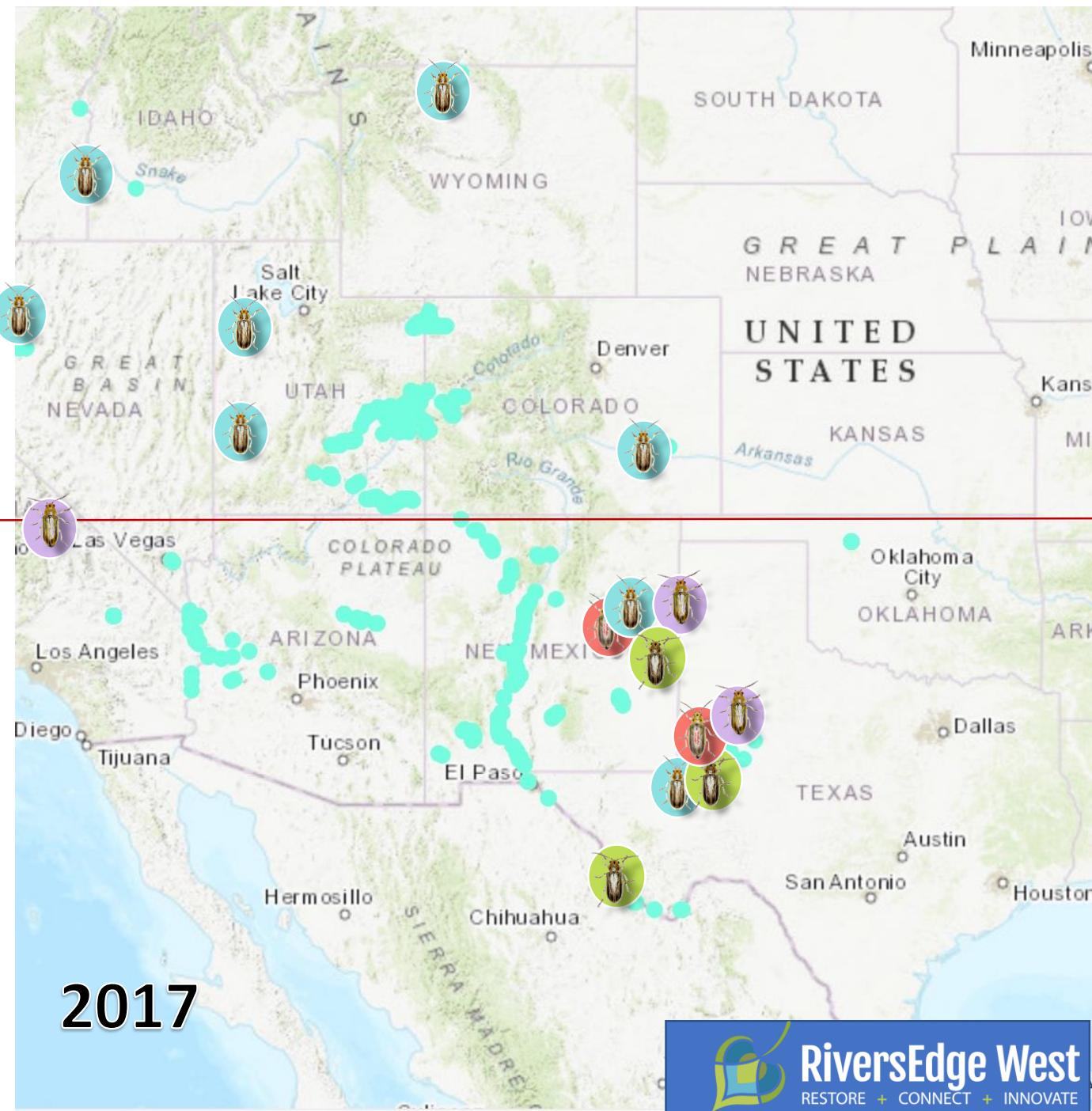
D. carinata



D. carinulata



Adapted from Bean, Kazmer, Gardner, Thompson, Reynolds, Keller, Gaskin (2013).



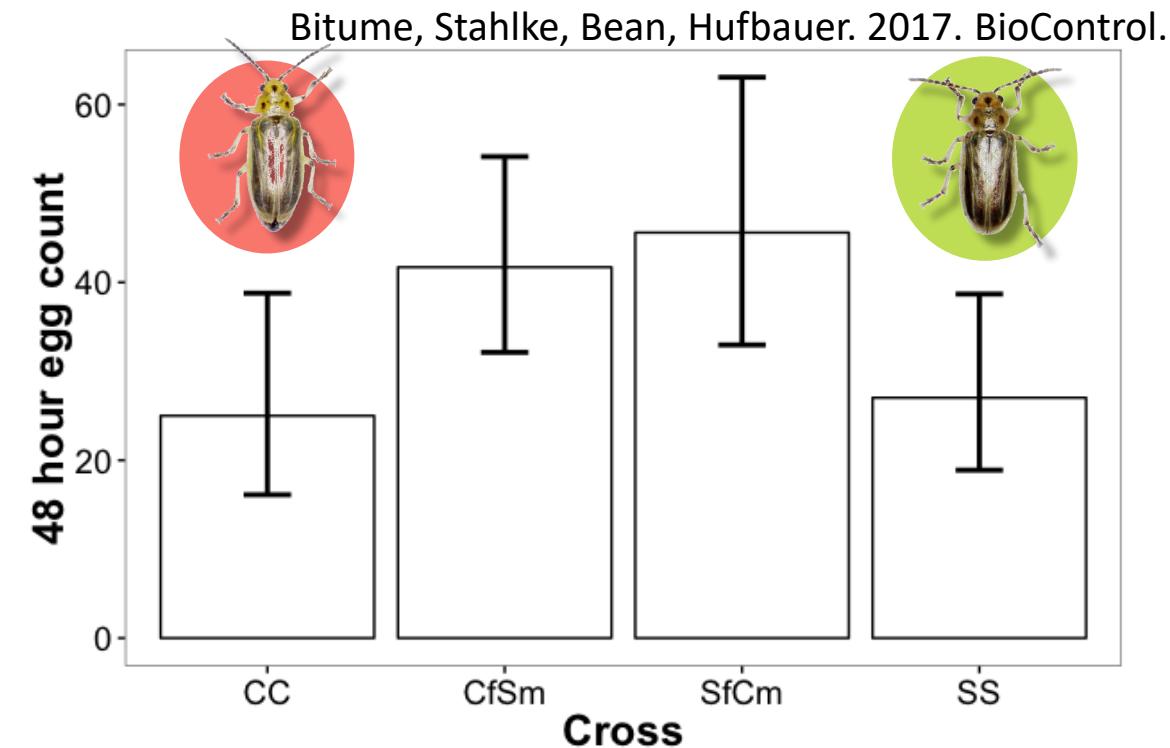
Where are these historically isolated lineages now?

Will range expansion and species interactions impact safety or efficacy?

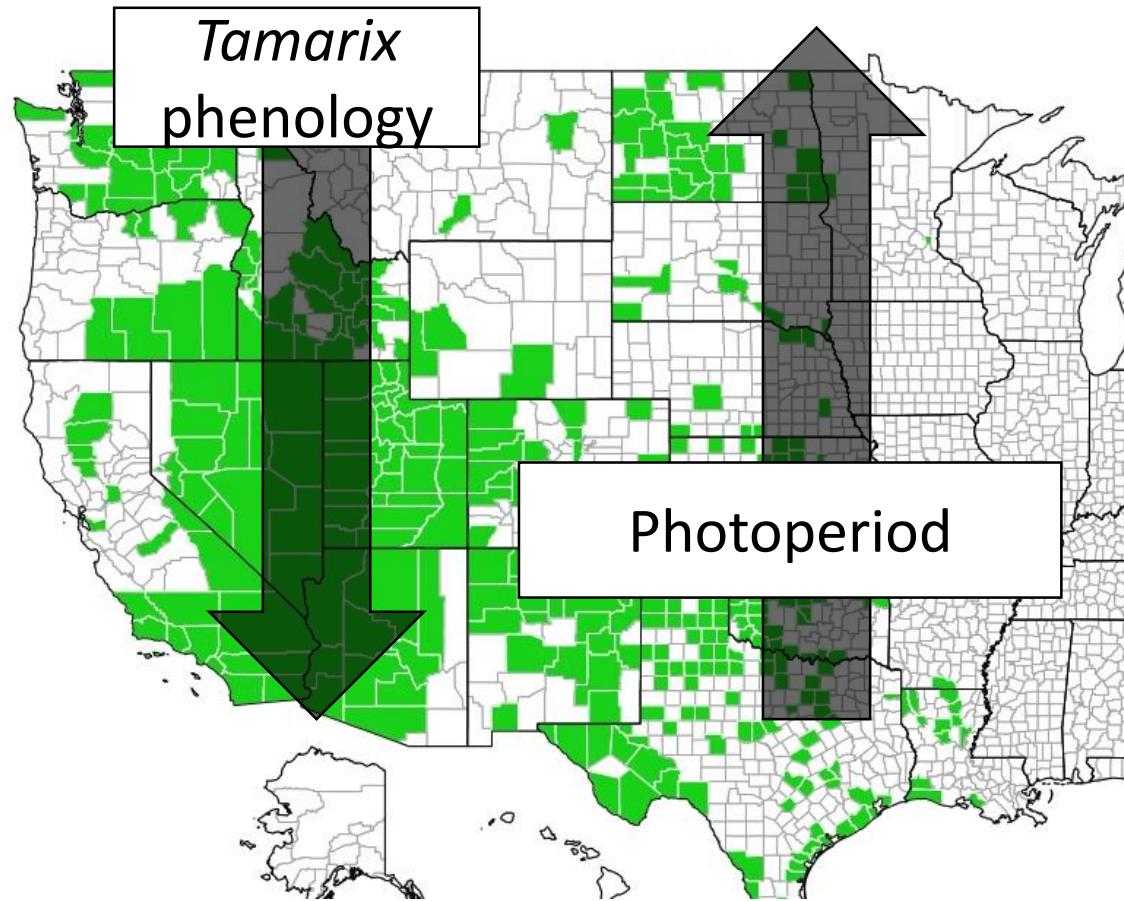
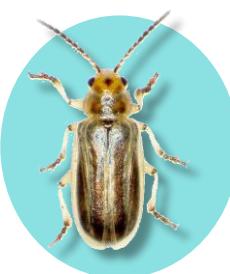
How can we use genetics/genomics to track and predict *Diorhabda* spp?

Hybridization may affect **safety** and **efficacy** in biological control

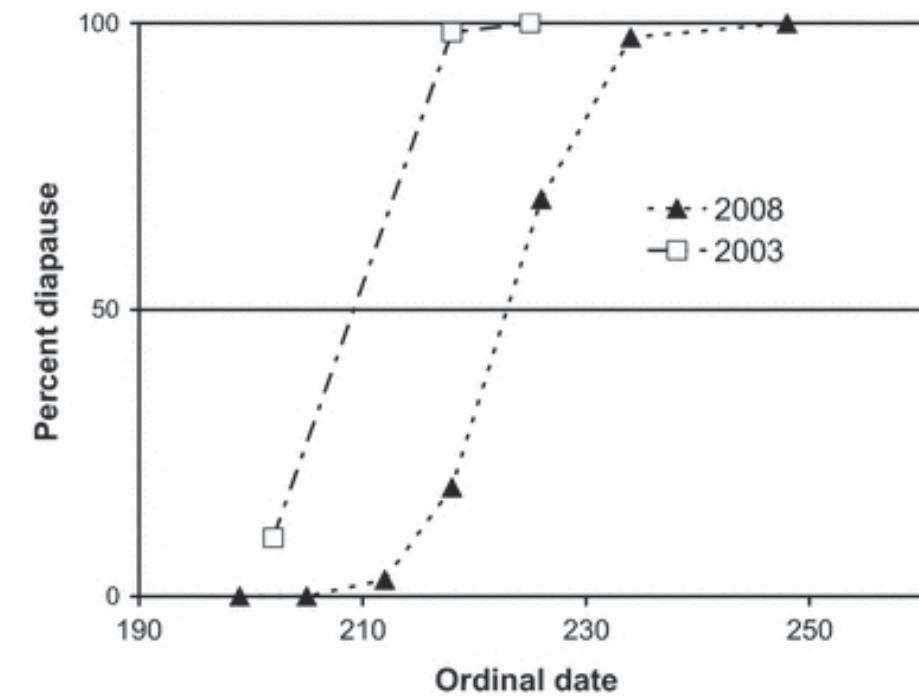
- Big boost in genetic diversity may increase likelihood of establishment.
- Novel genetic combinations in a novel environment could yield transgressive phenotypes.
- Hybrids could be sterile, ineffective



Range expansion enabled in part by evolution in diapause response.



Rapid evolution of phenology

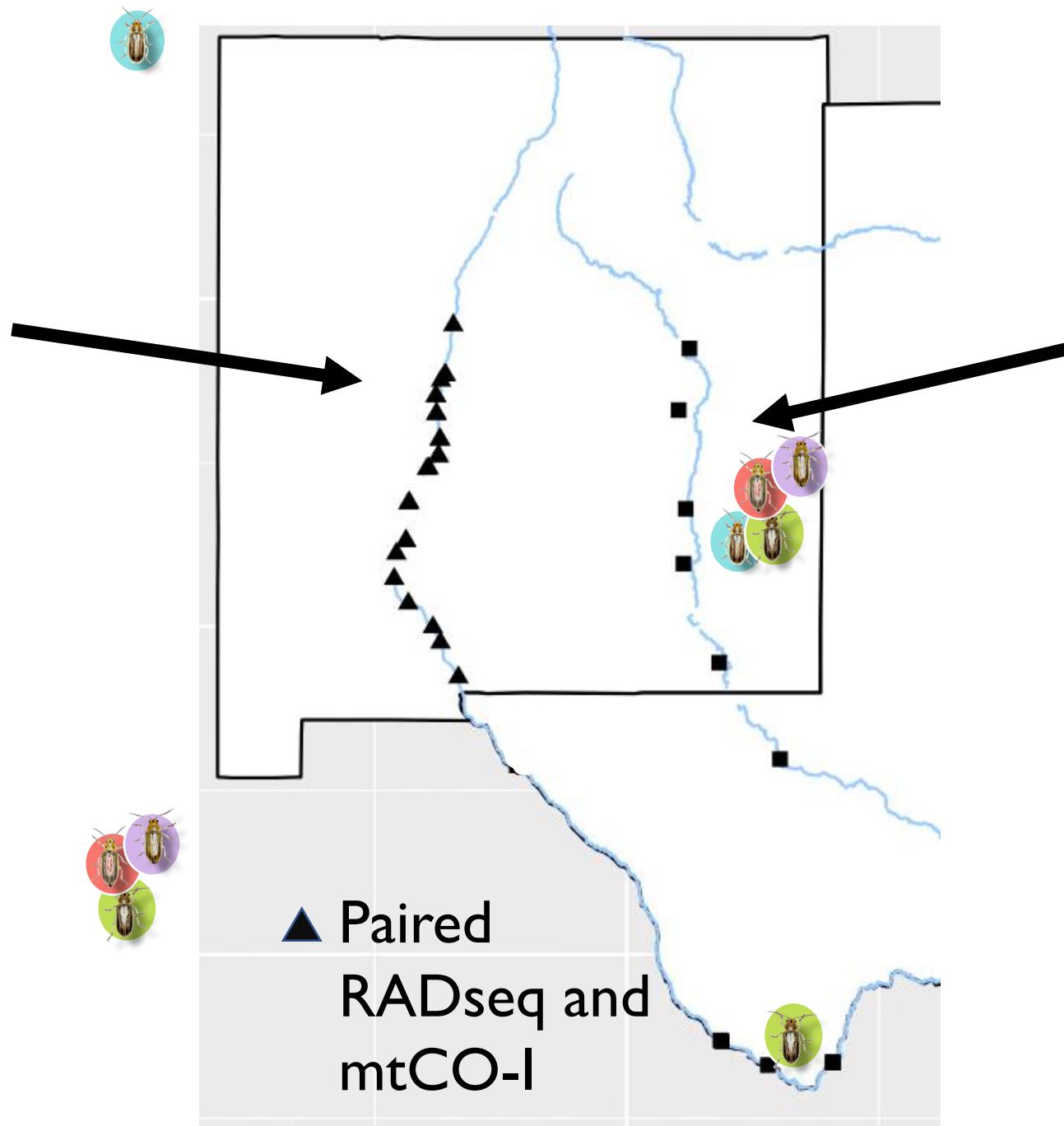


Bean, Dalin, Dudley. 2012. Evo. App.

adapted from Hultine, Bean, Dudley, Gehring. (2015).

Rio Grande

Tracking population dynamics over a full season



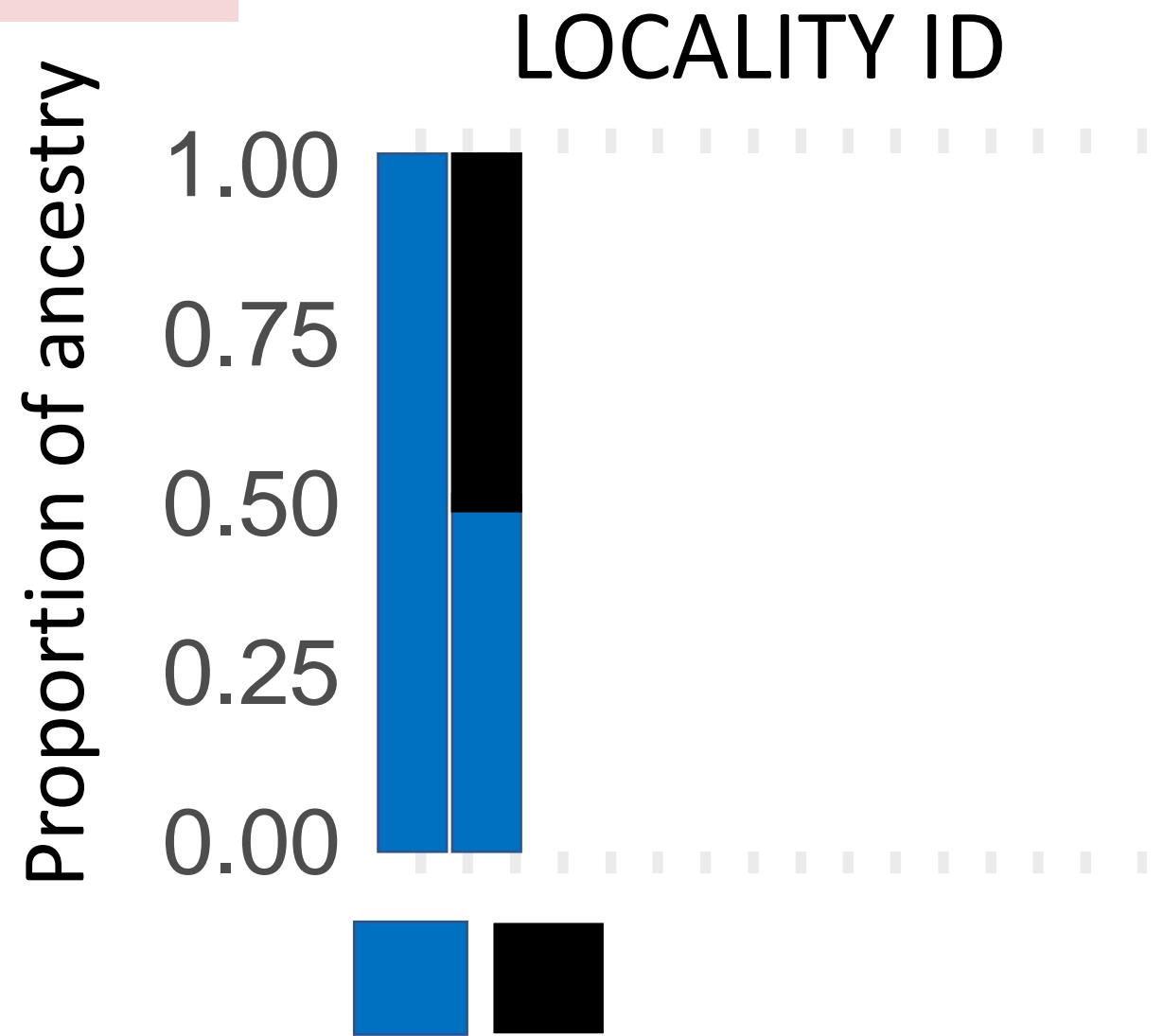
Pecos River

Part of a large project to broadly characterize hybridization and population structure

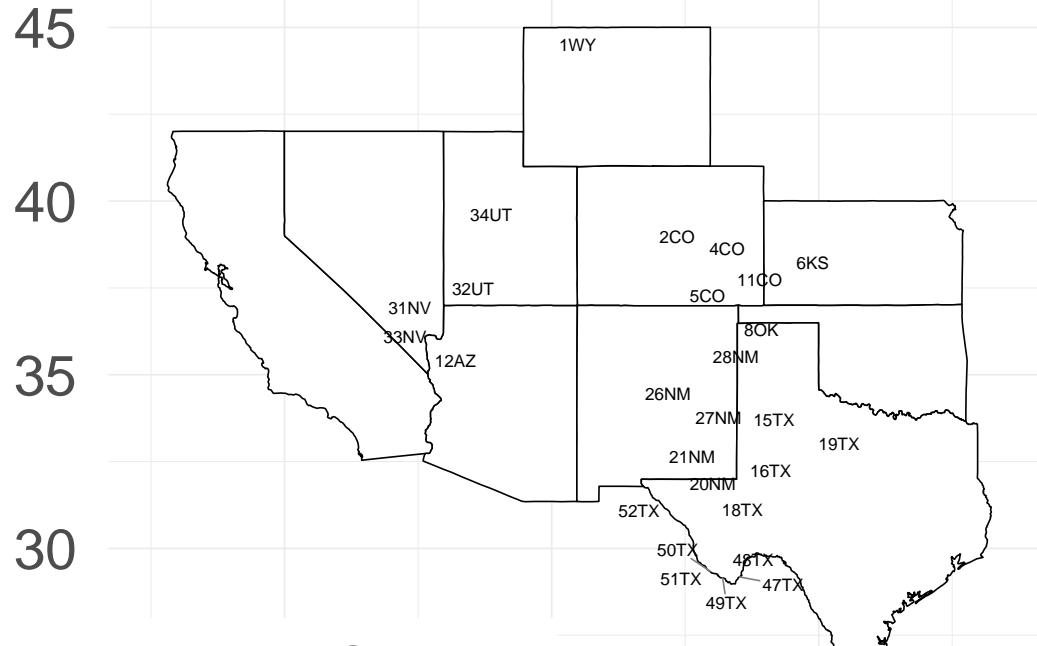
Genomic ancestry assignment

RADseq: Fast, relatively
cheap, flexible,
molecular method to
assess **genomic**
variation
9,125 SNPs

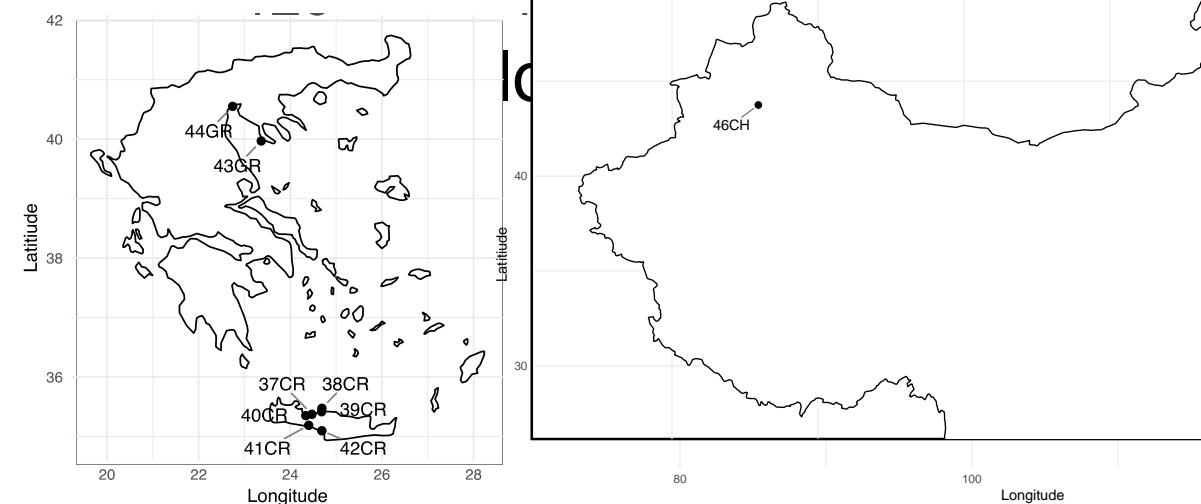
STRUCTURE, a Bayesian
clustering algorithm
Probability(X|K)



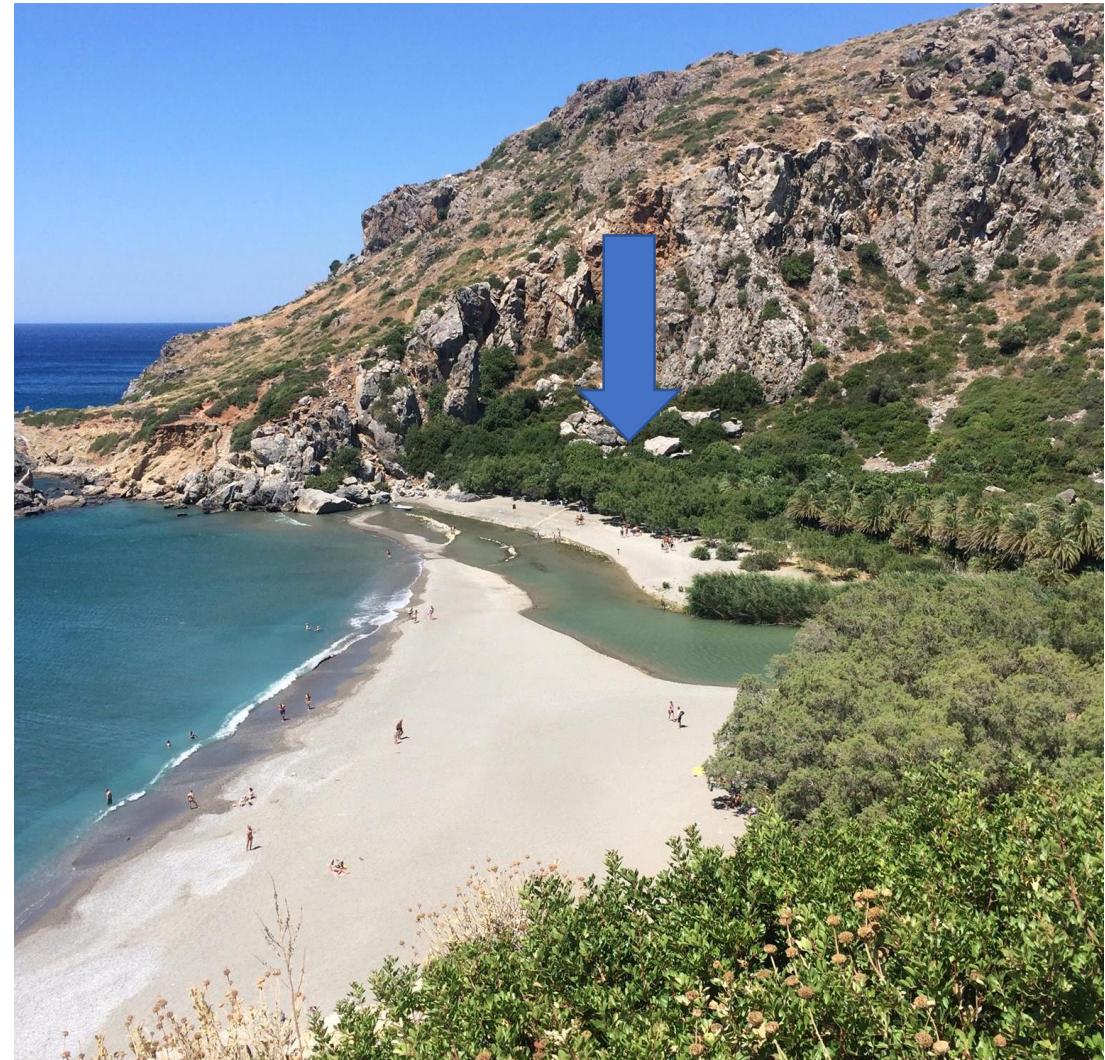
Southwest United States



Mainland Greece
& Crete

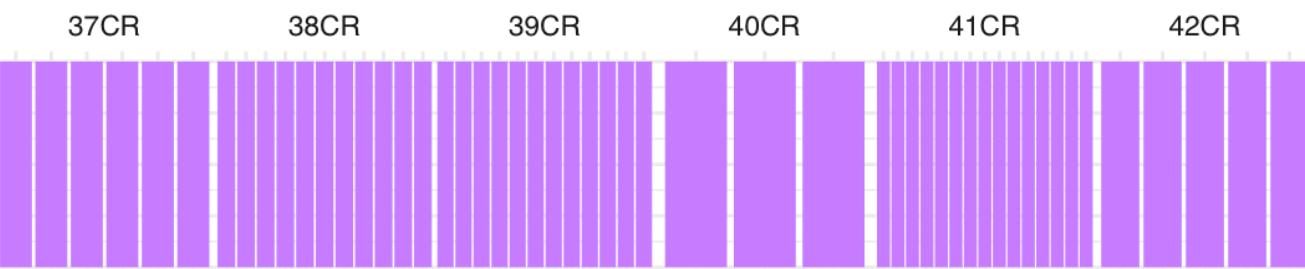
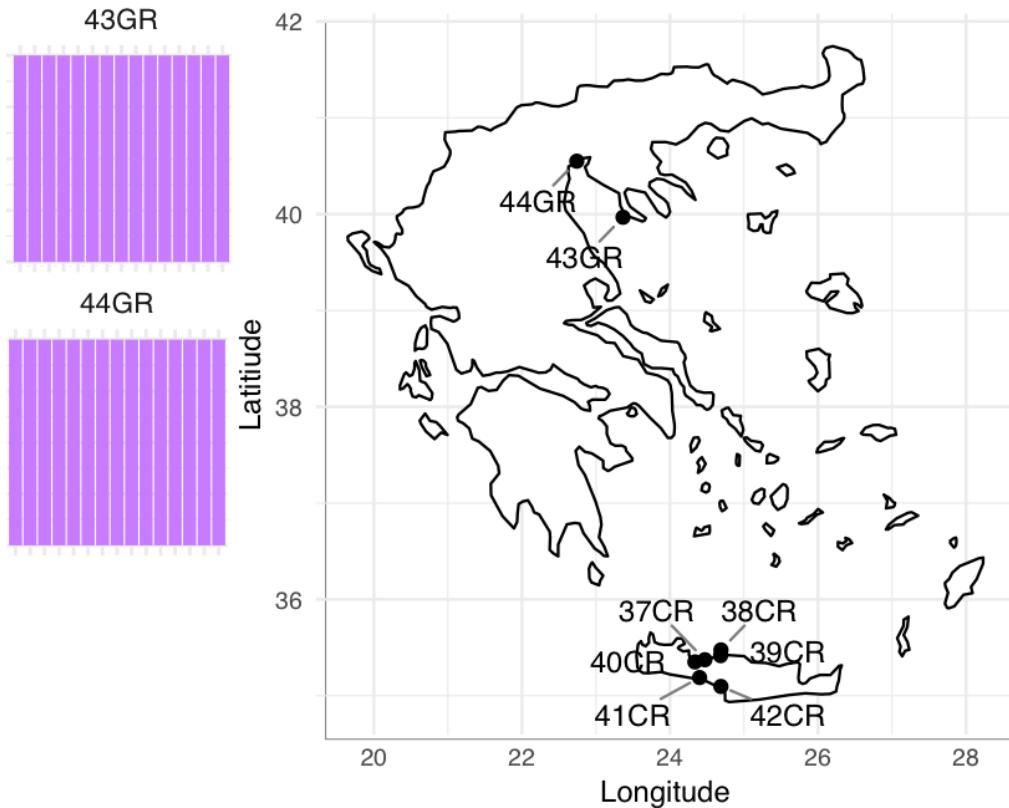


Collected field samples from lab cultures and over 50 sites within introduced and native range

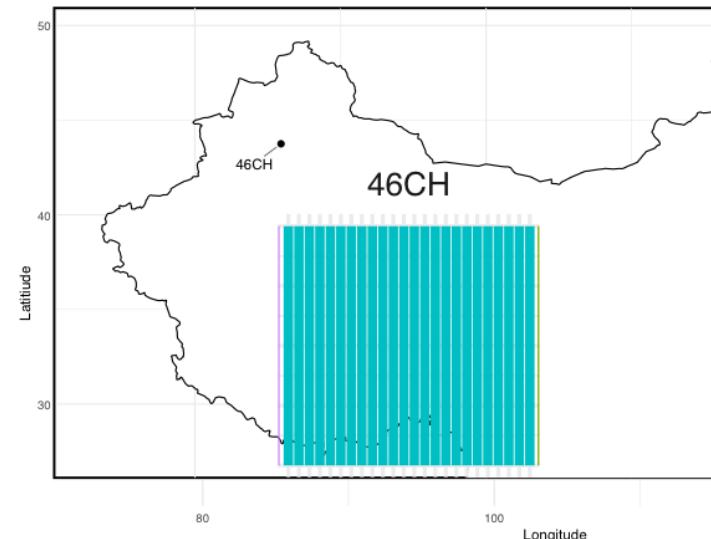


No hybridization was detected in the native range or lab.

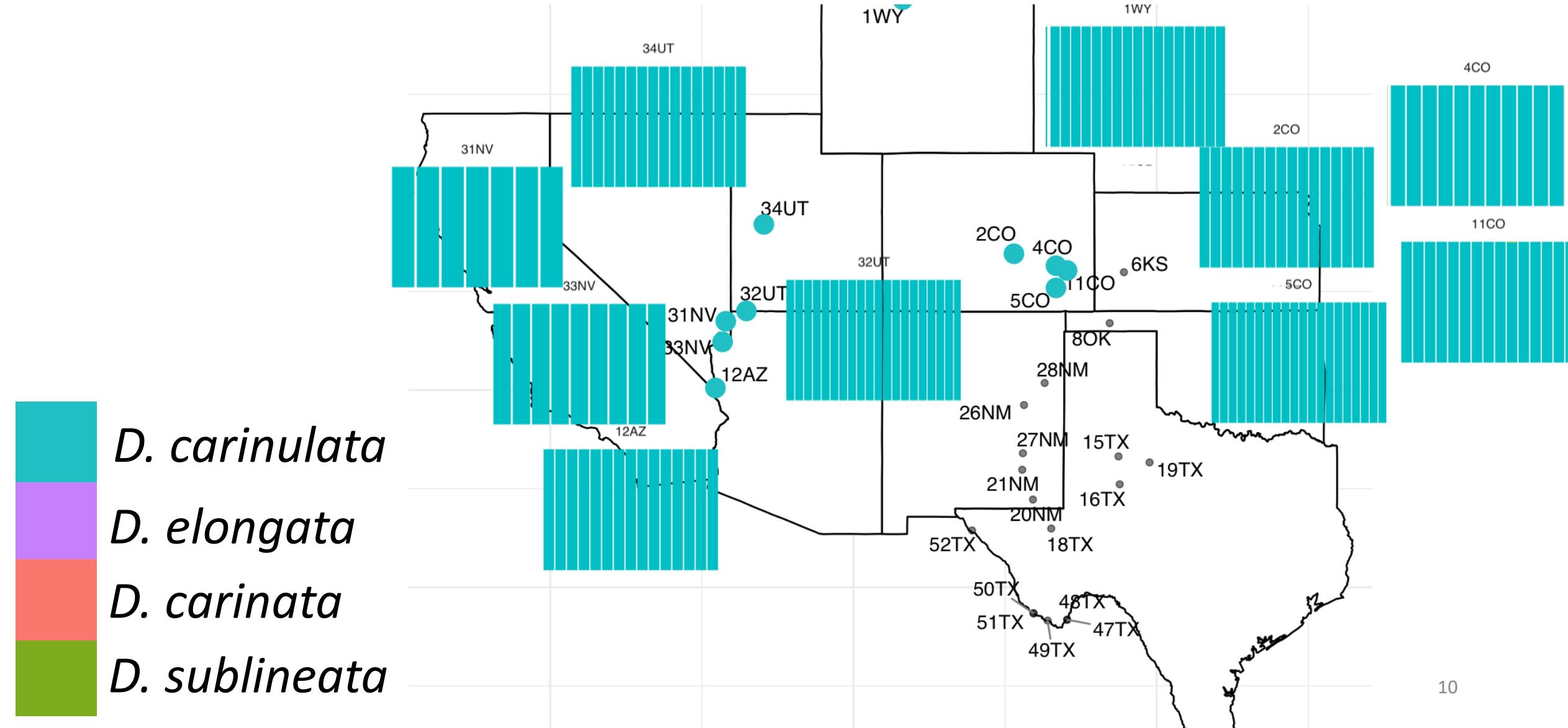
Mainland Greece & Crete



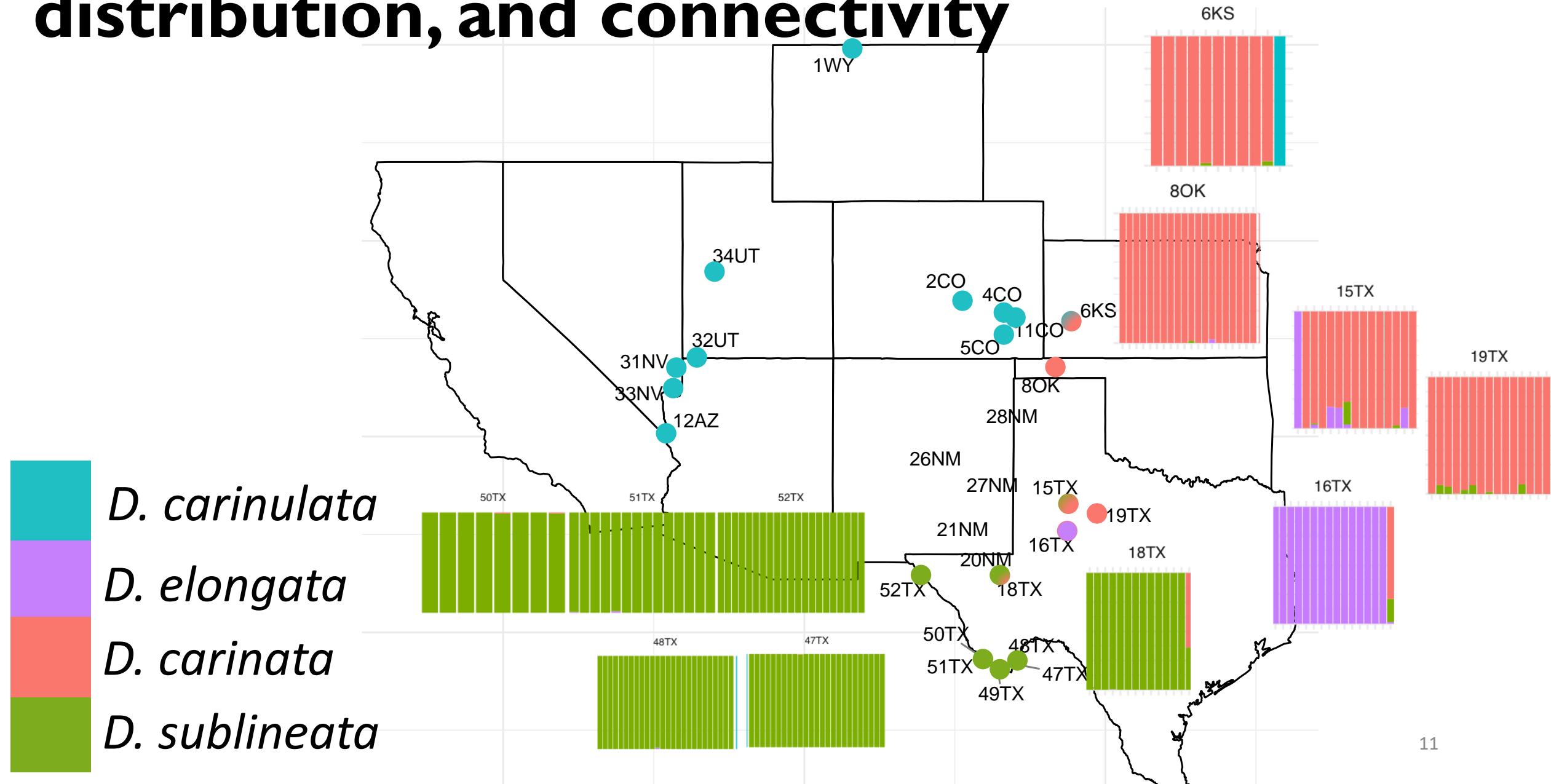
Xinjiang, China



Ancestry assignment reflects introduction, re-distribution, and range expansion

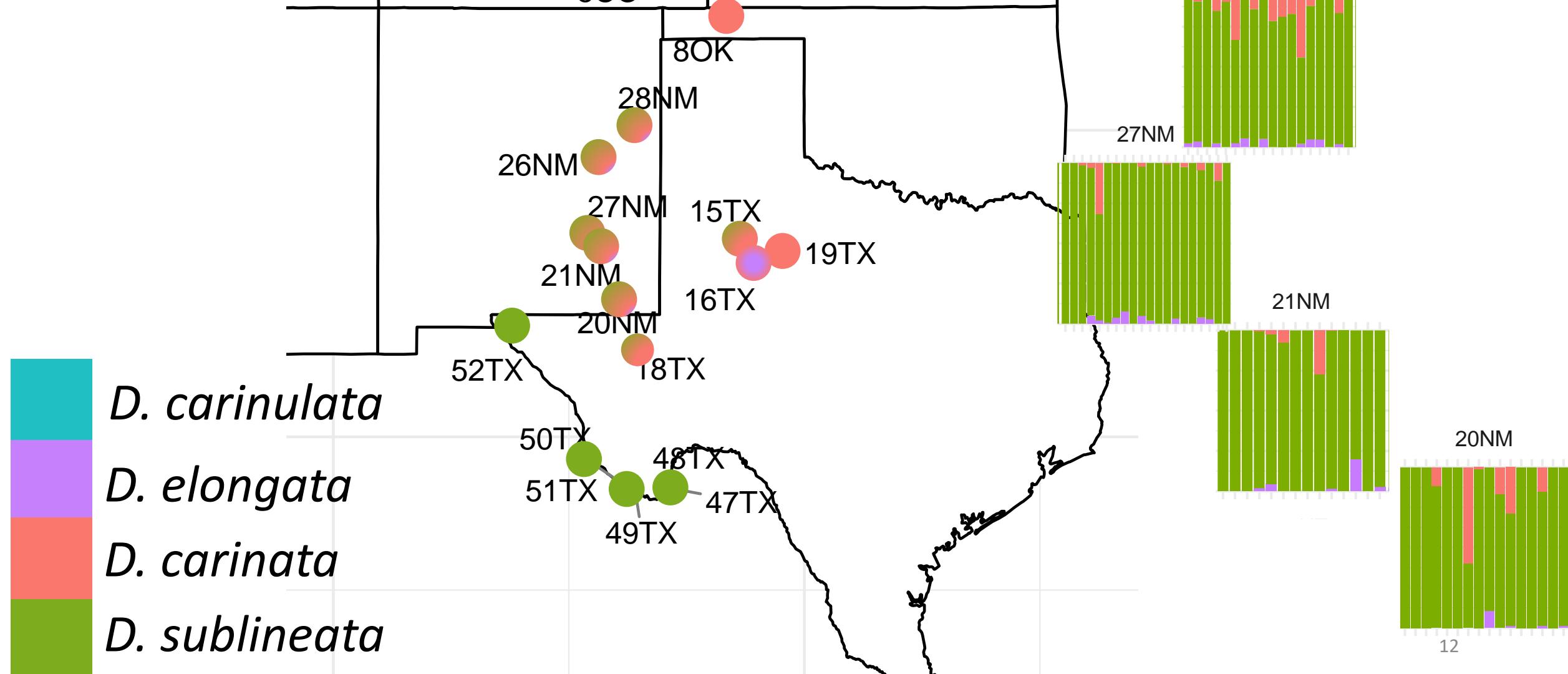


Ancestry assignment reflects introduction, redistribution, and connectivity



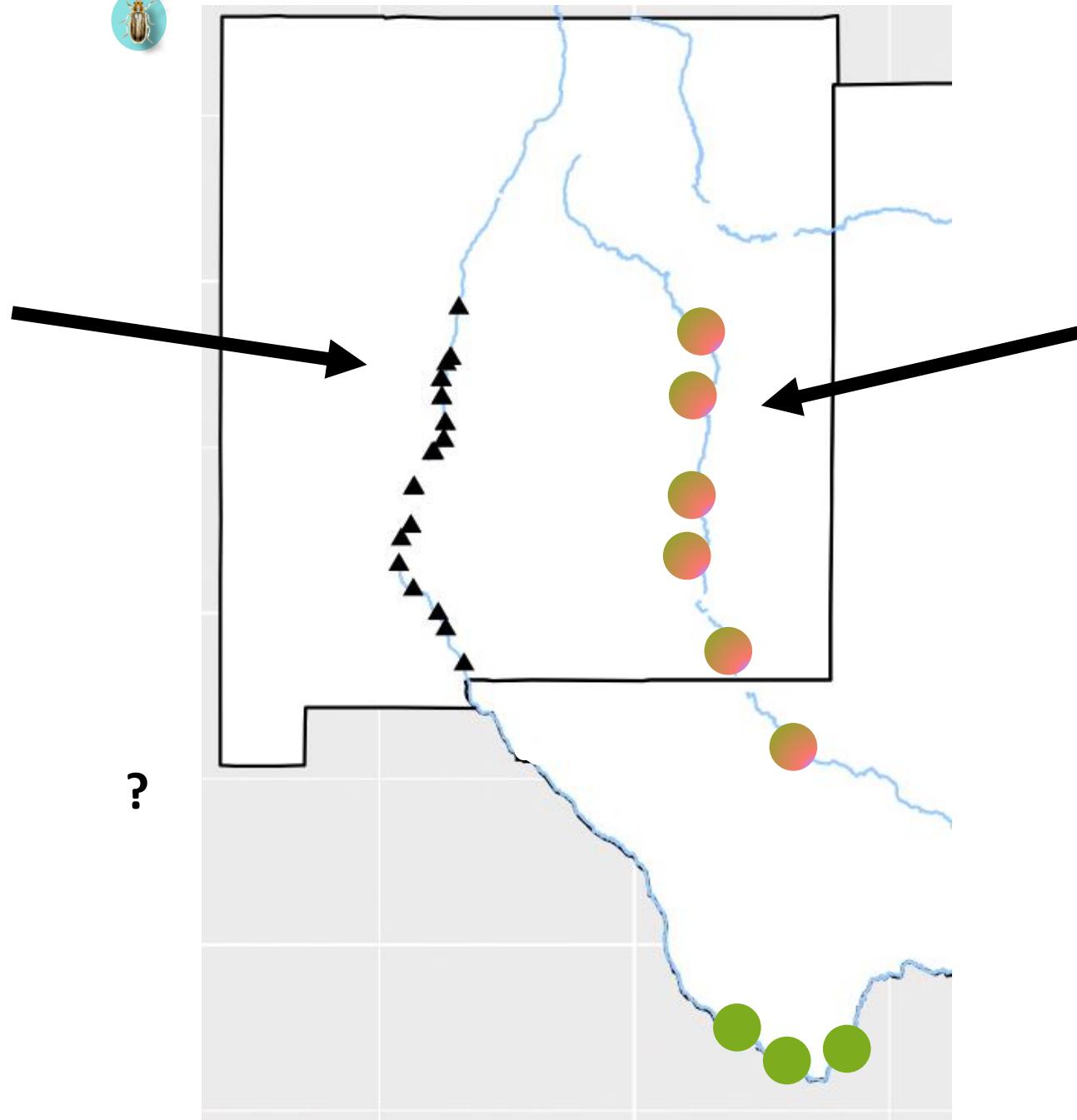
28NM

Ancestry assignment reveals widespread hybridization at contact zones



Rio Grande

Tracking population dynamics over a full season



Pecos River

If present will hybridize
Pure populations still existed on

Rio Grande

- Part of a focused effort to track range expansion and species interactions (Levi 2:00)
- 5 samples per site in May, July, and September
- Performed RADseq & sequenced mitochondrial marker COI

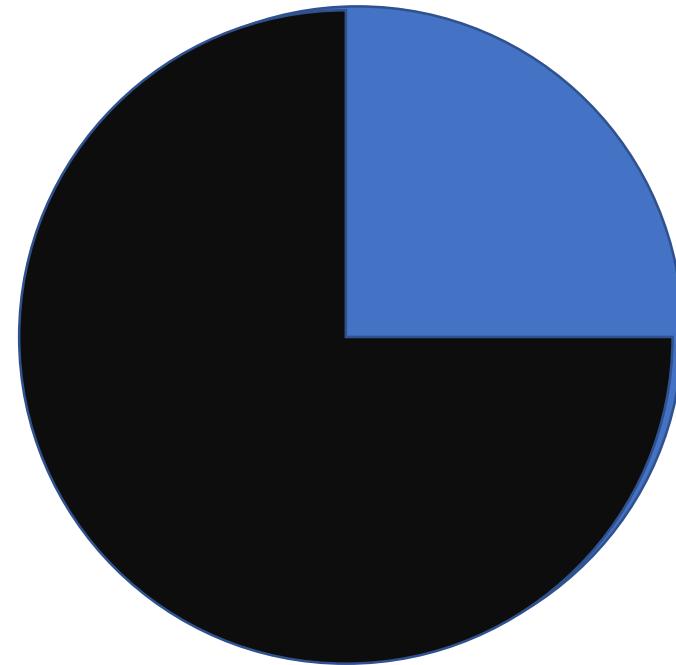
CO-I ancestry assignment

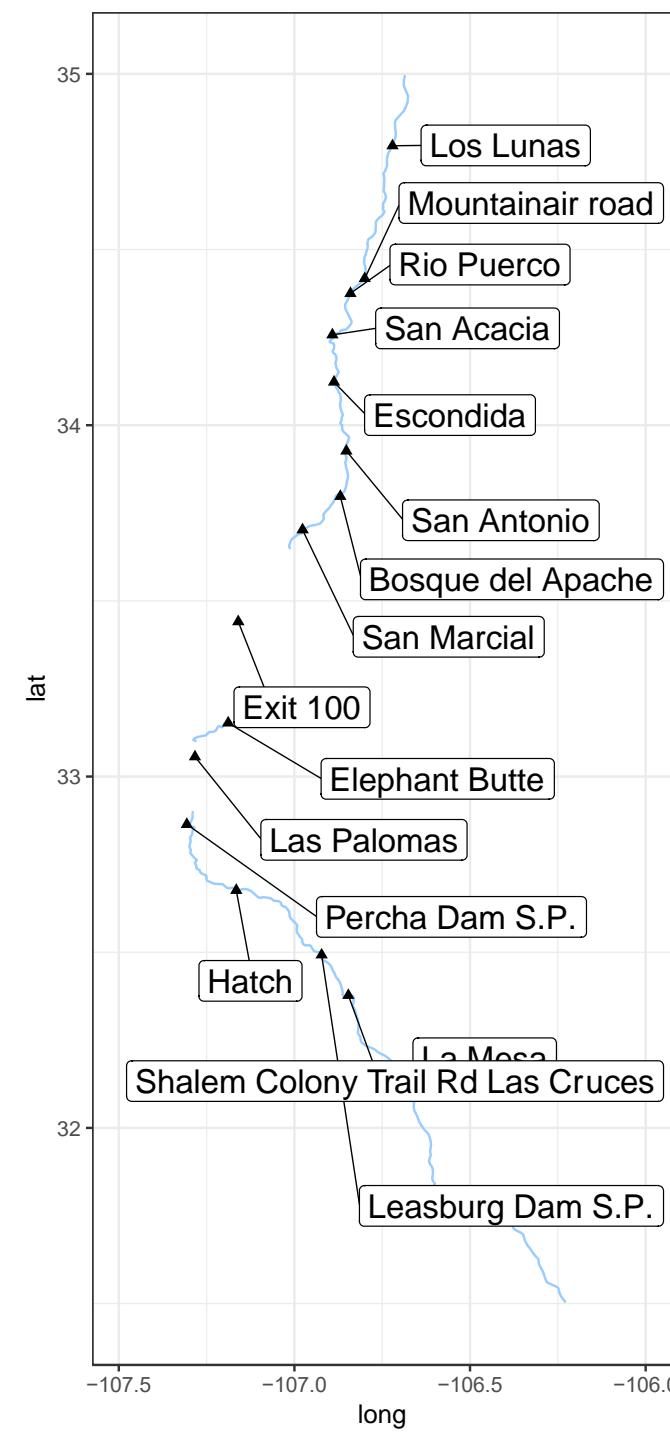
Mitochondrial marker
Cytochrome Oxidase subunit-I
1296 bp Sanger sequencing

Can handle more degraded samples, single samples at a time

98% sequence homology with morphologically IDed samples
(Bean et al 2013)

LOCALITY ID

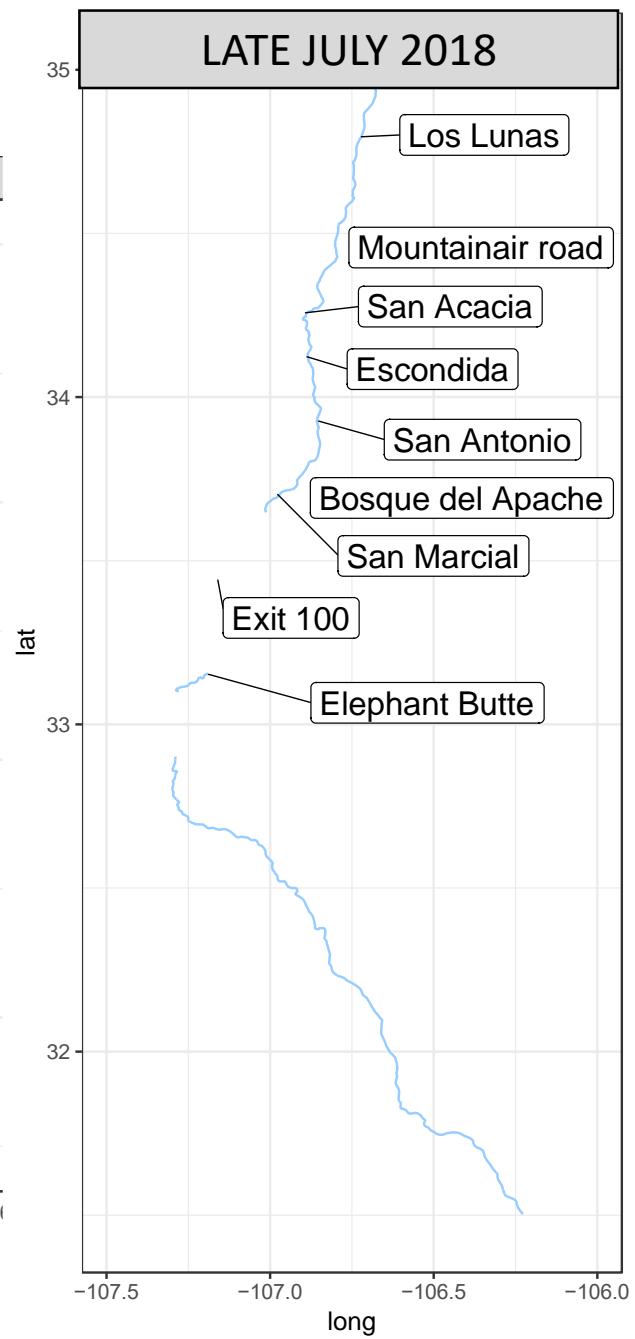




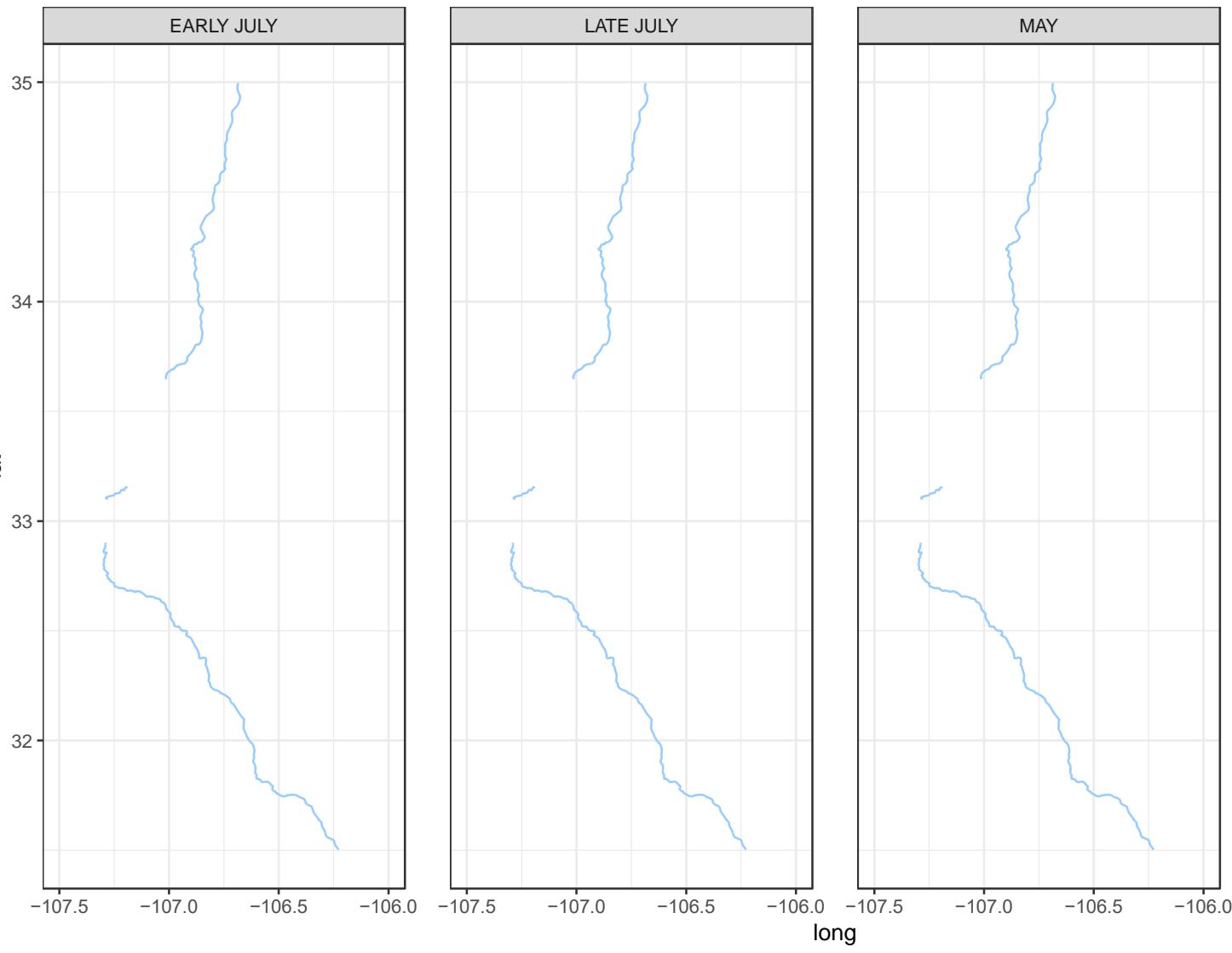
CO-I haplotype

long
carinulata elongata sublineata

LATE JULY 2018



2017



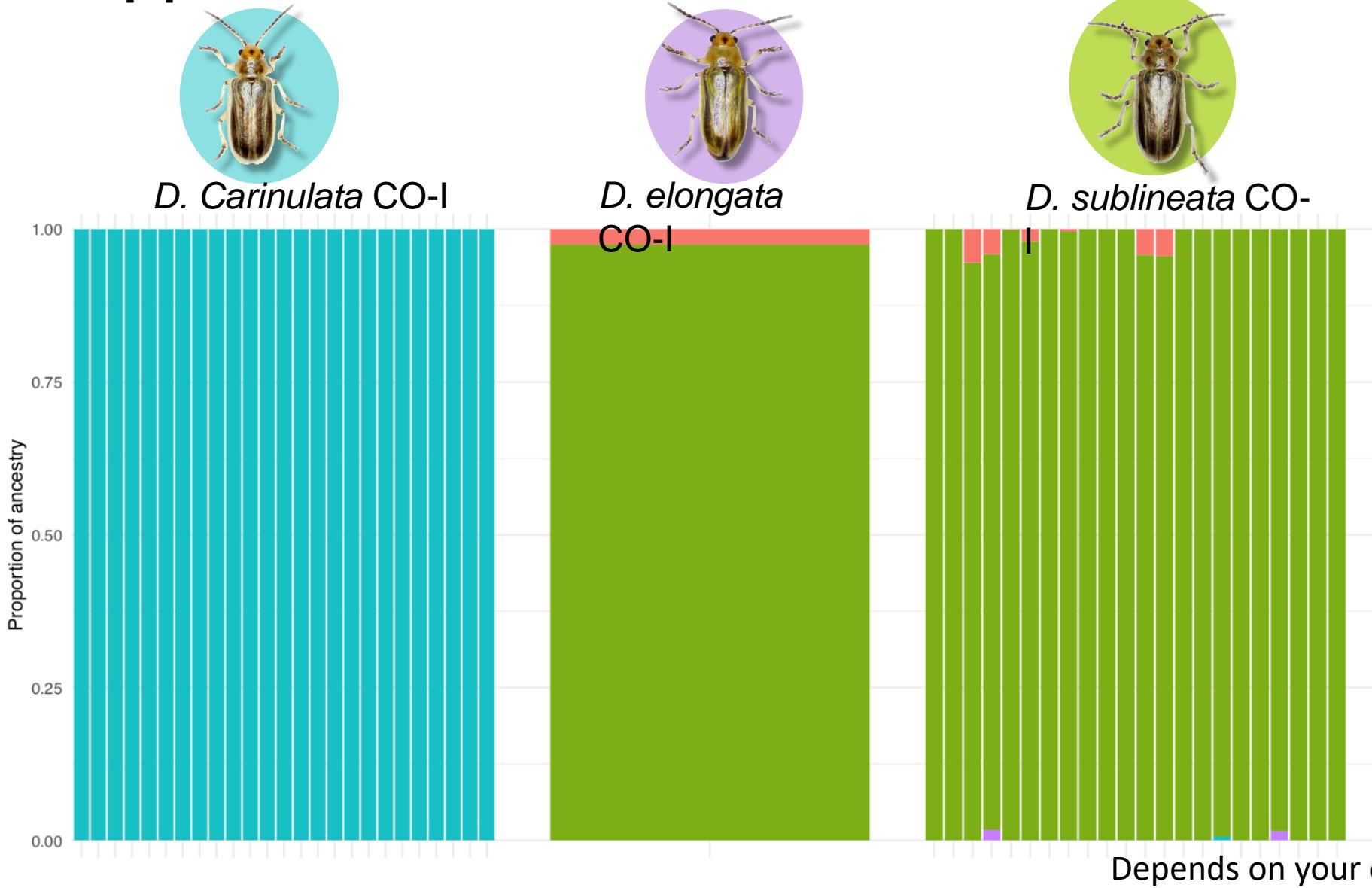
CO-I haplotype

carinulata

elongata

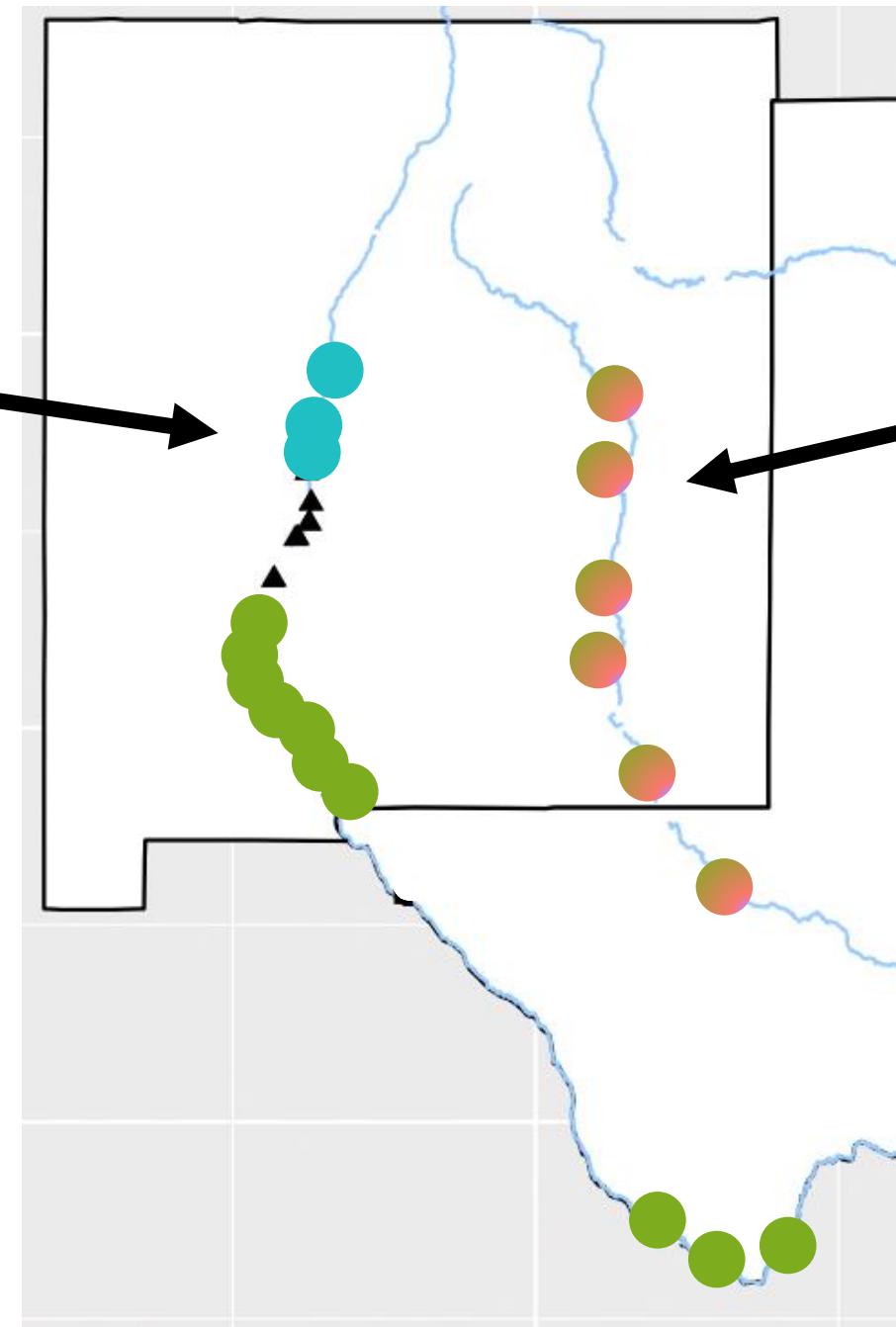
sublineata

What's the best way to track *Diorhabda* spp?



Rio Grande

- Population composition changed rapidly over a season,
- But competition could slow range expansion
- CO-I remains useful for rapid ID

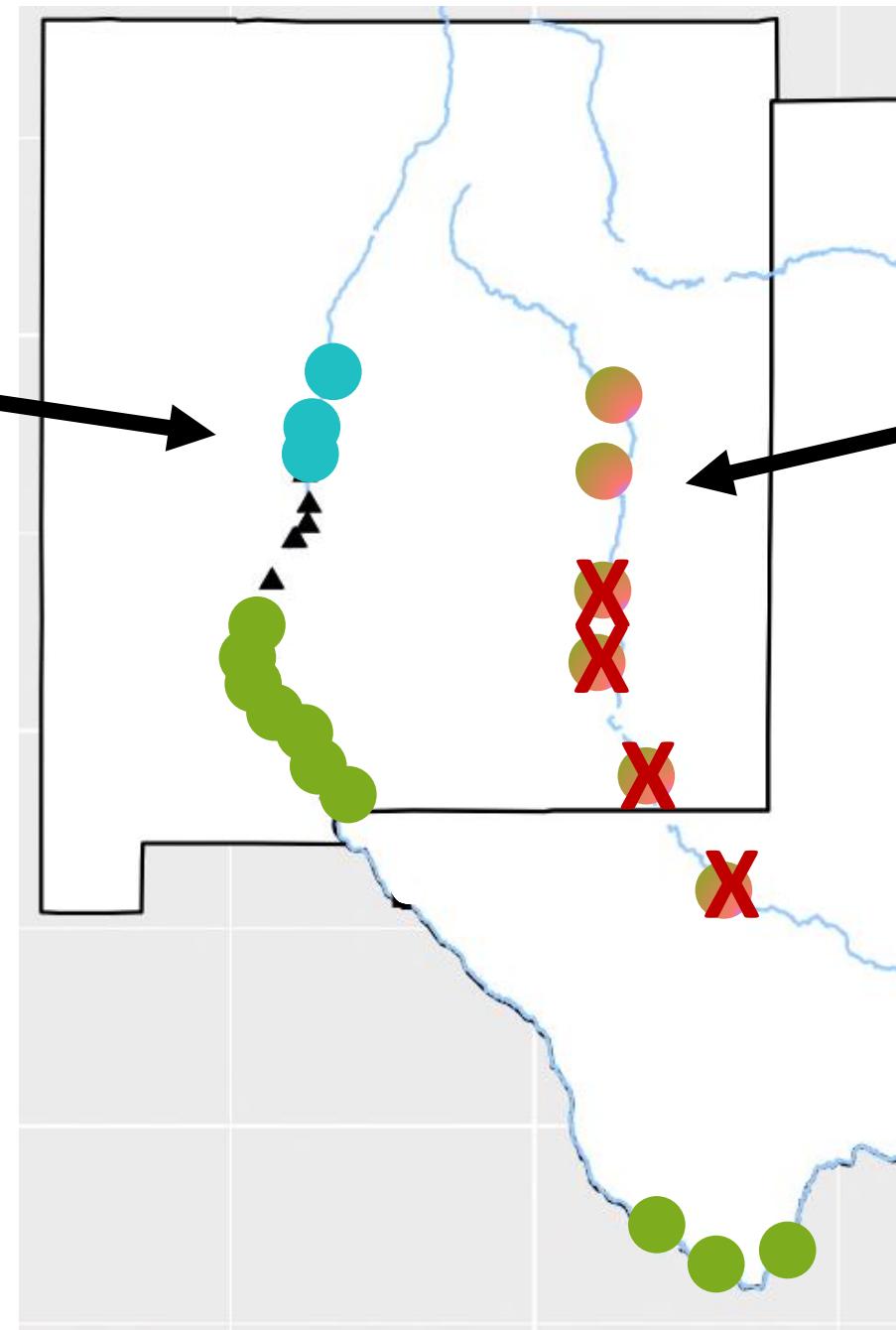


Pecos River

- If present, will hybridize
- Lack of hybrids elsewhere: strong role for connectivity? possible role for maladaptation?

Rio Grande

- Population composition changed rapidly over a season,
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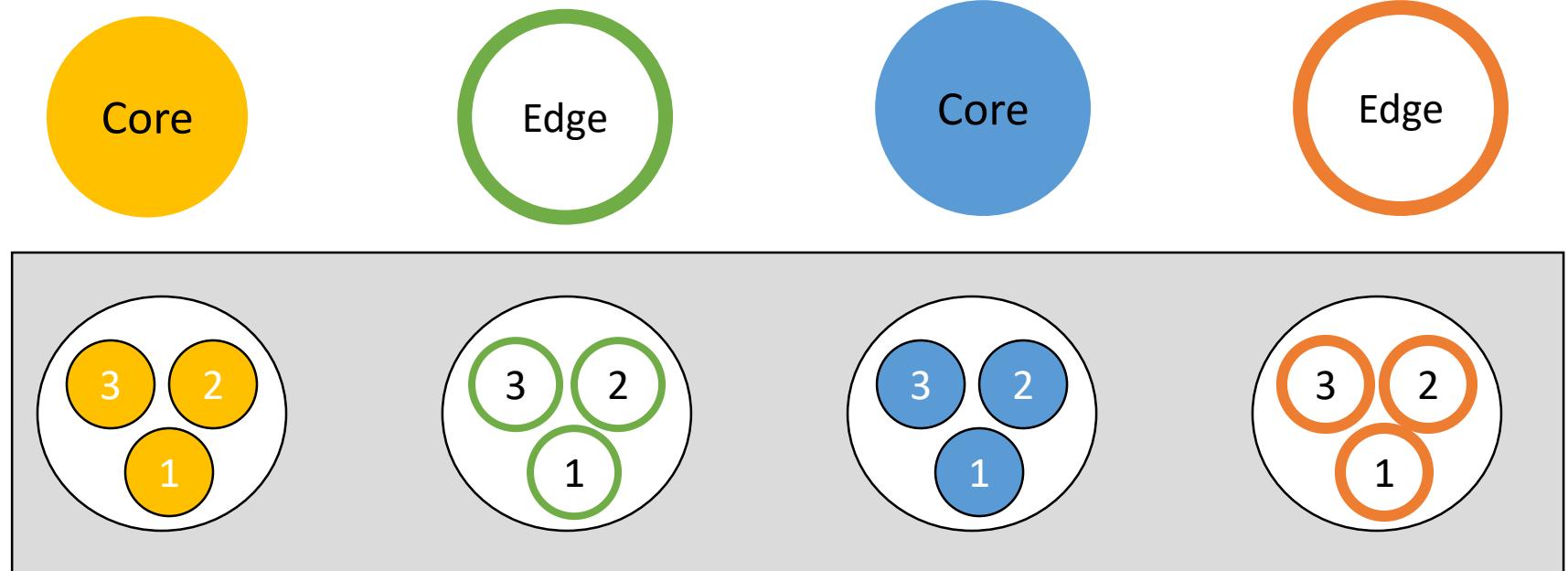


Pecos River

- If present, will hybridize
- Lack of hybrids elsewhere: strong role for connectivity? possible role for maladaptation?

Many hybrid populations seem to have disappeared

Phenotyping



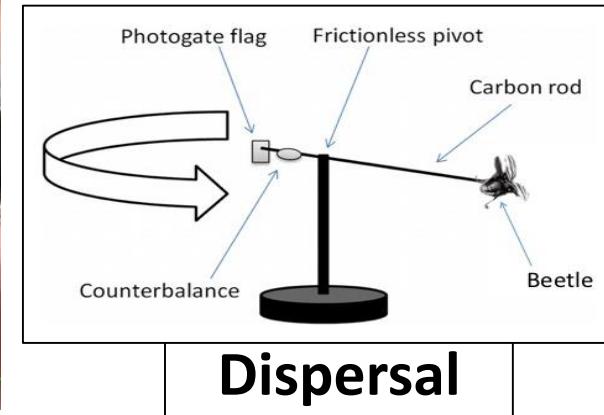
♀ — ♂



Diapause



Fecundity



Dispersal



Host choice

Acknowledgments

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René Sforza

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Karen Rosen

Nina Louden

Sonya Daly

Beyond

Hohenlohe Lab

Tom Dudley

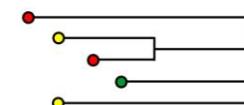
James Tracy

Massimo Cristofaro

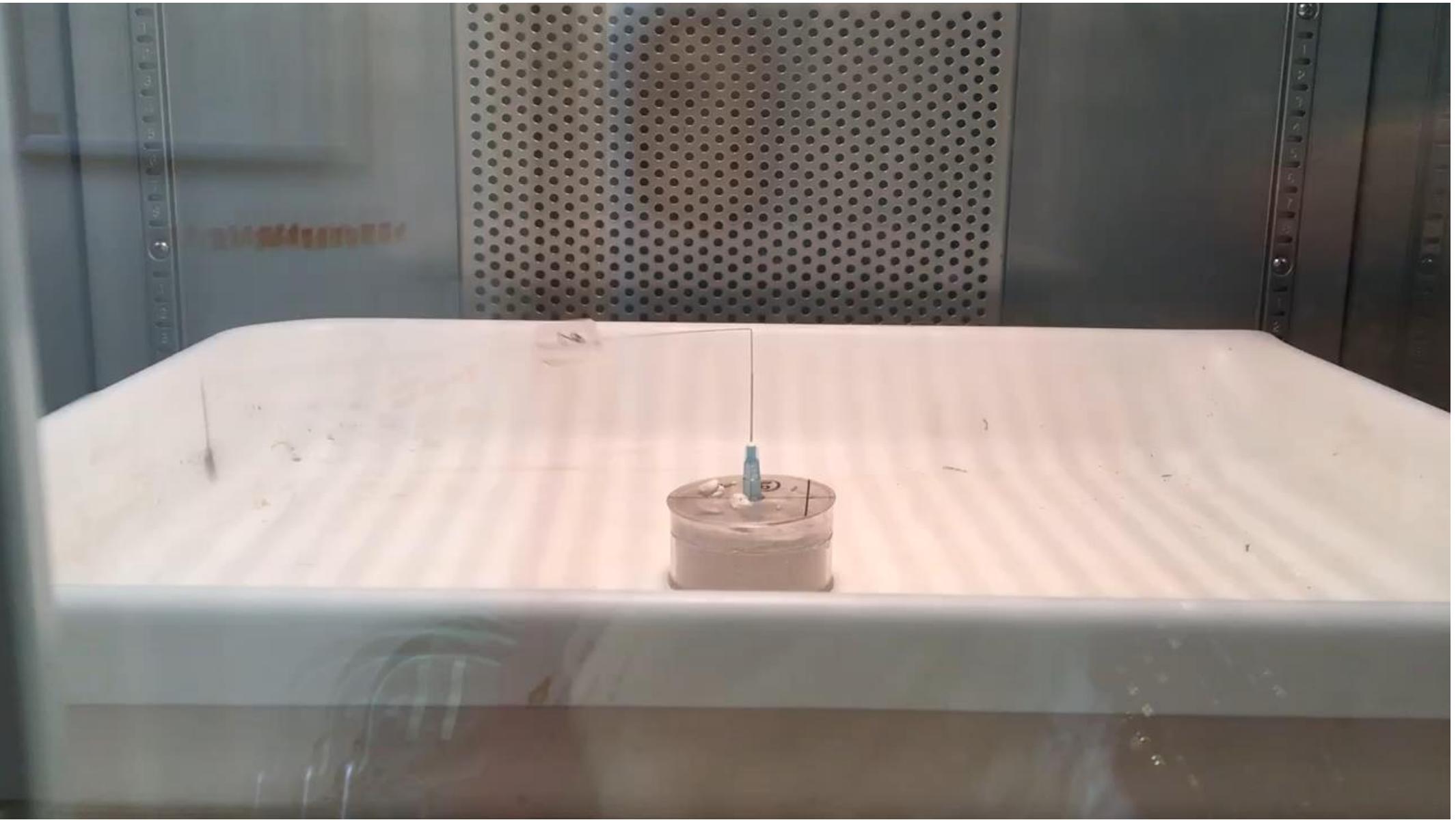


United States Department of Agriculture
National Institute of Food and Agriculture

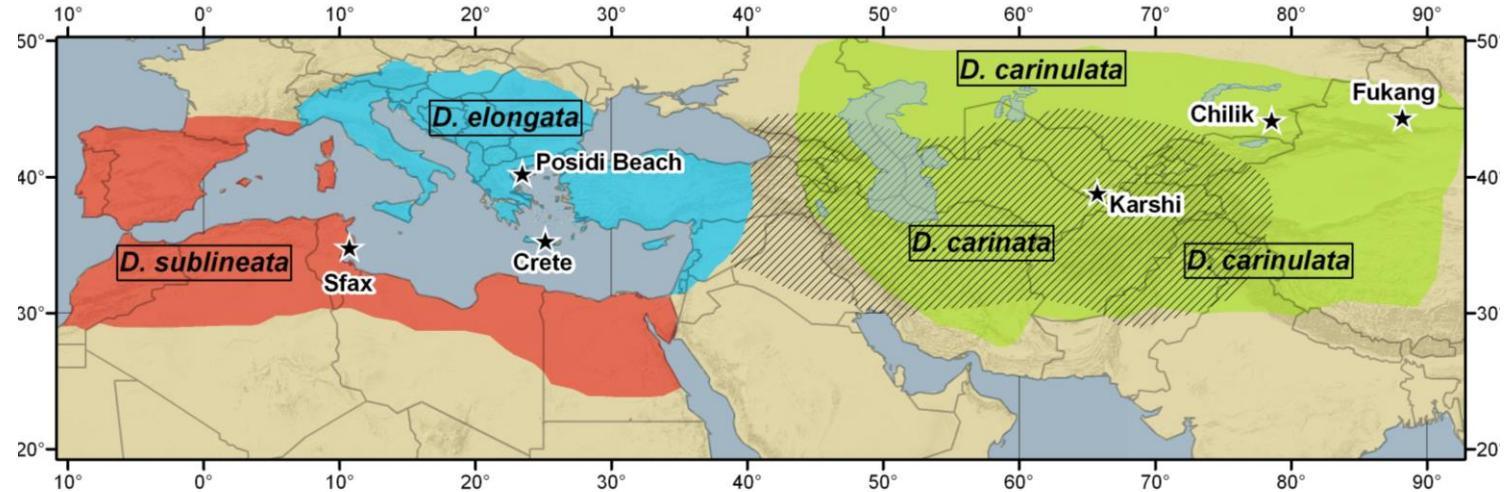
USDA G-13528

 **ibest**

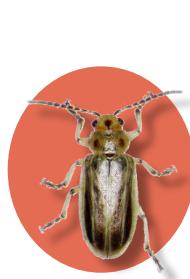
The logo for ibest features the letters "ibest" in a large, bold, black sans-serif font. To the left of the text is a graphic element consisting of four horizontal lines of varying lengths, each ending in a colored circle (red, yellow, green, blue).



Thank you!



The Tamarisk Leaf Beetle, *Diorhabda* spp.



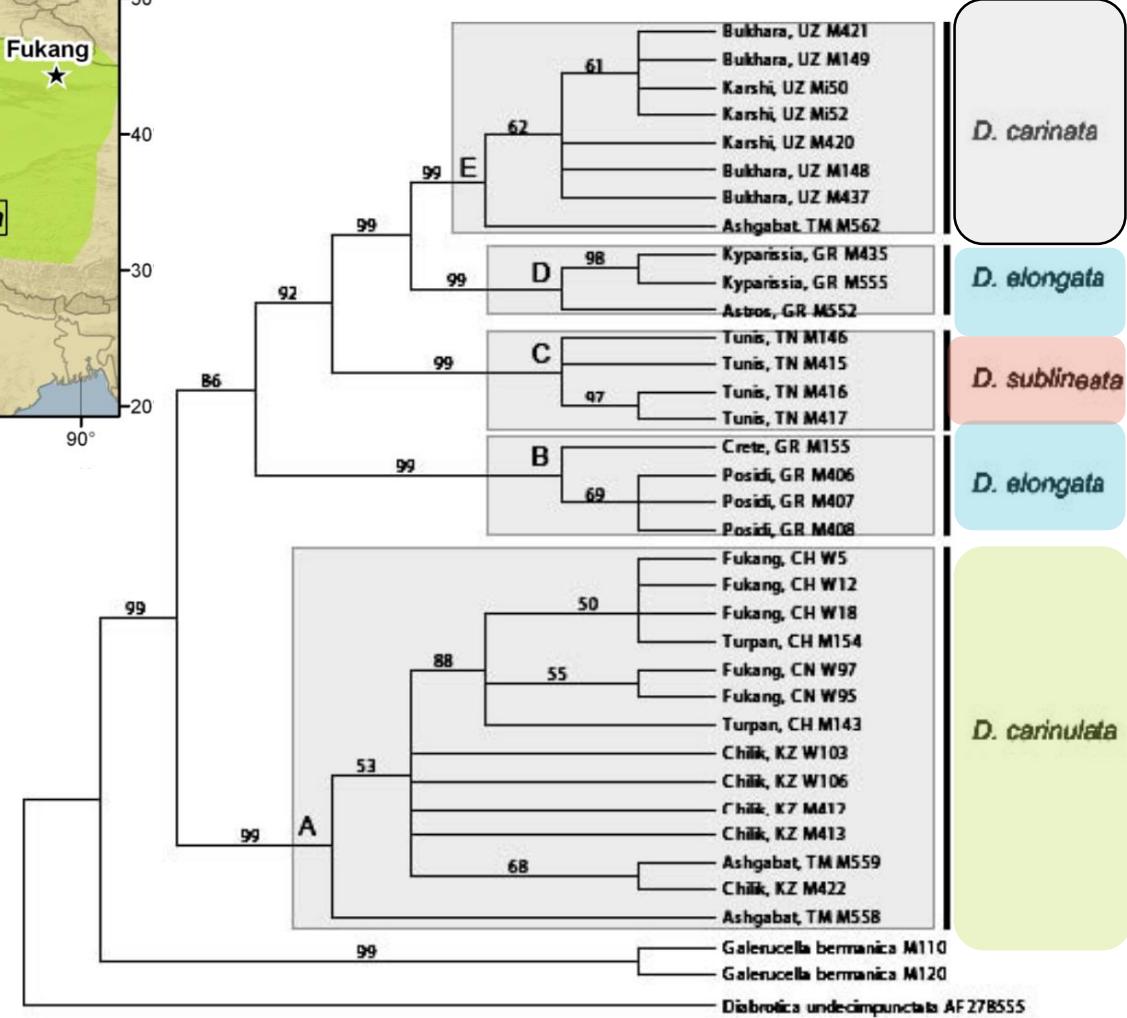
D. elongata



D. carinata

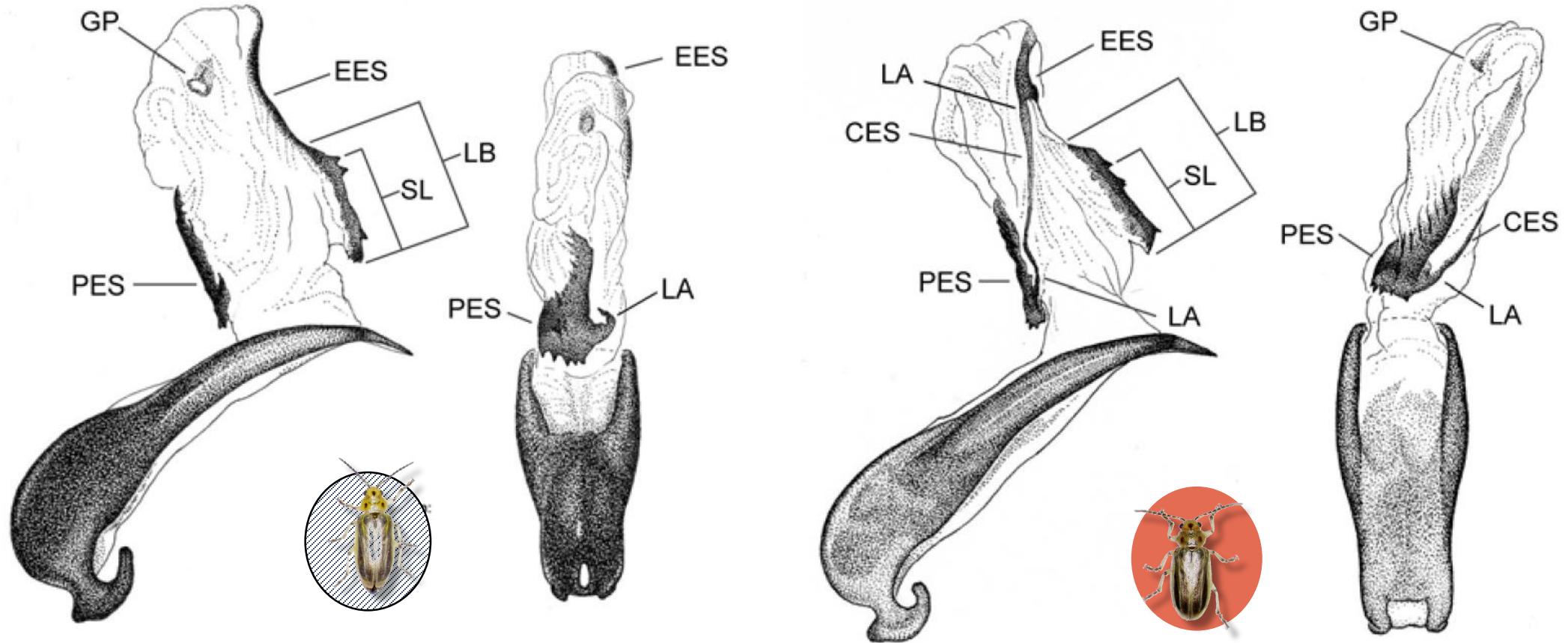


D. sublineata



Maximum parsimony 50% majority consensus mtCOI

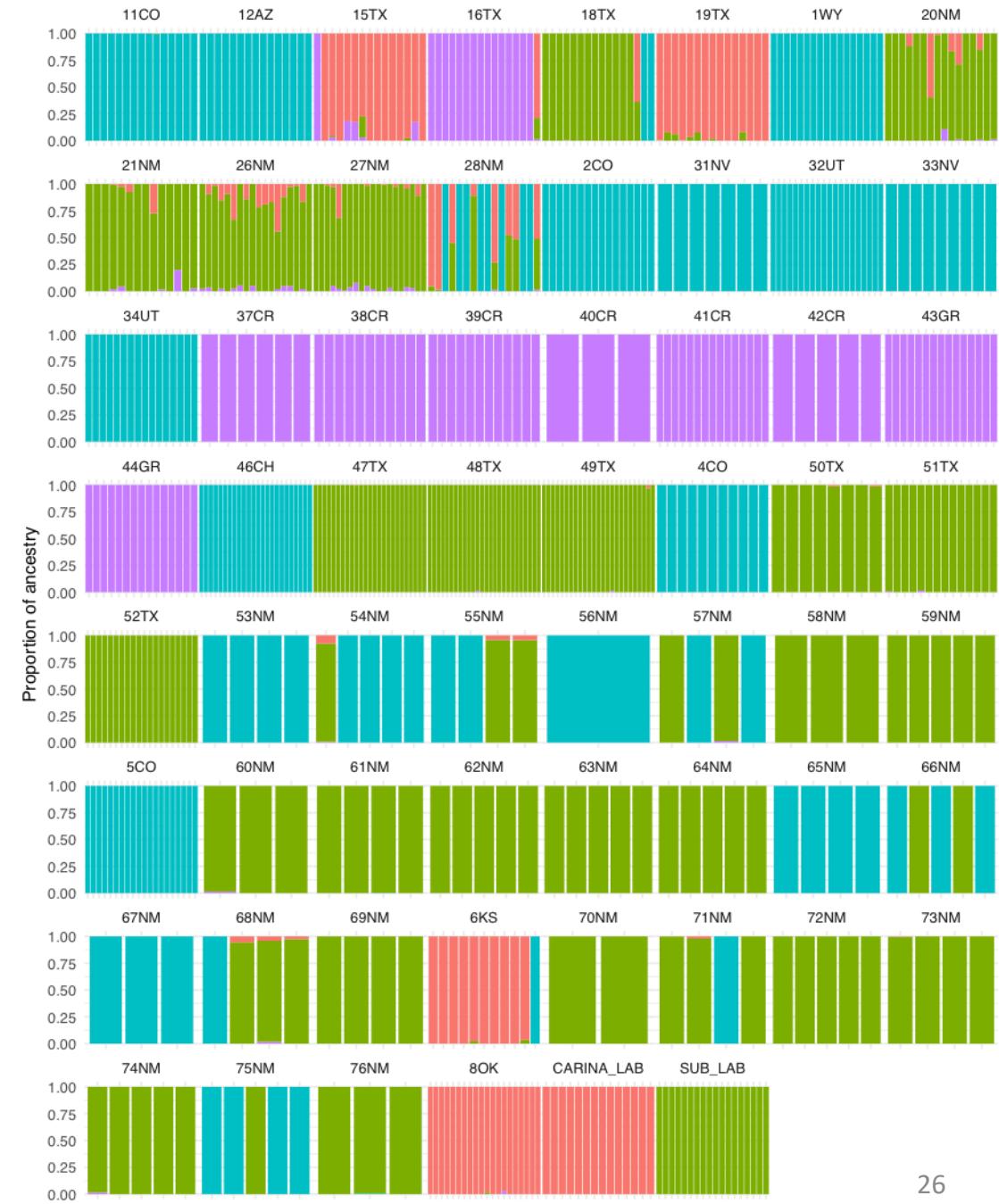
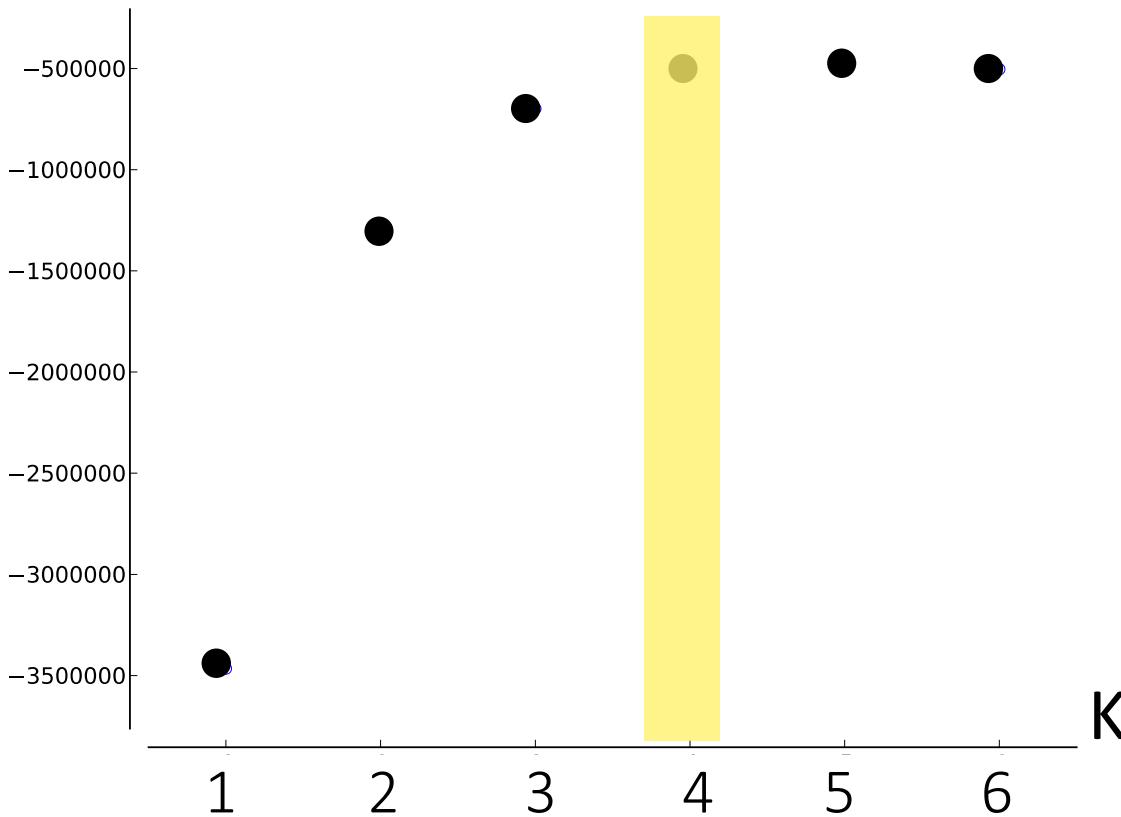
Internal morphology defines species delimitation and identification



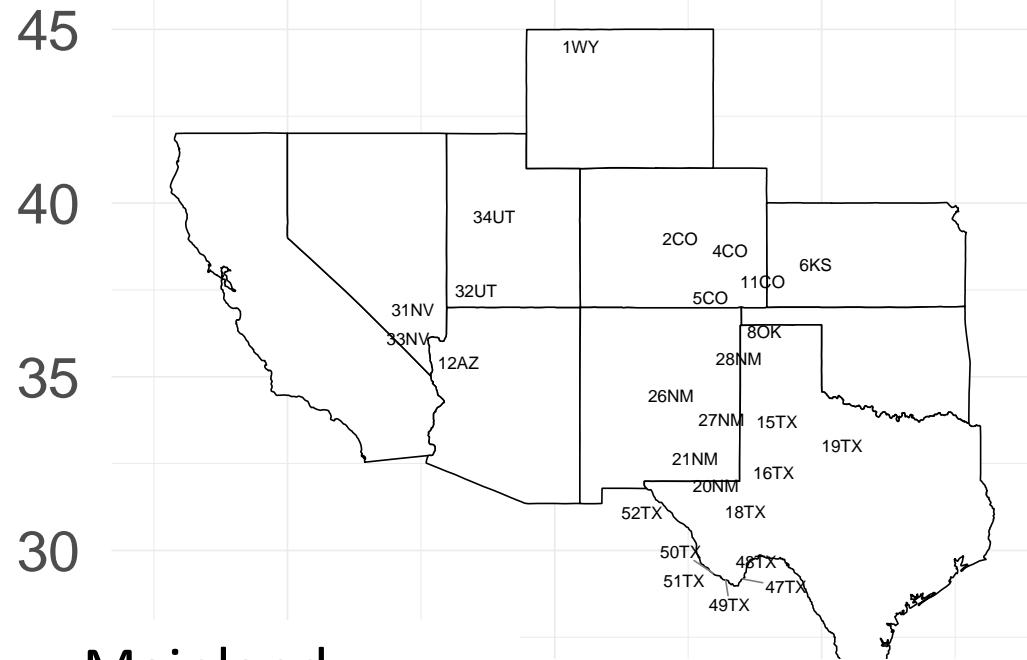
Tracy & Robbins. 2009. Zootaxa.

K=4 reflects species identity for known samples

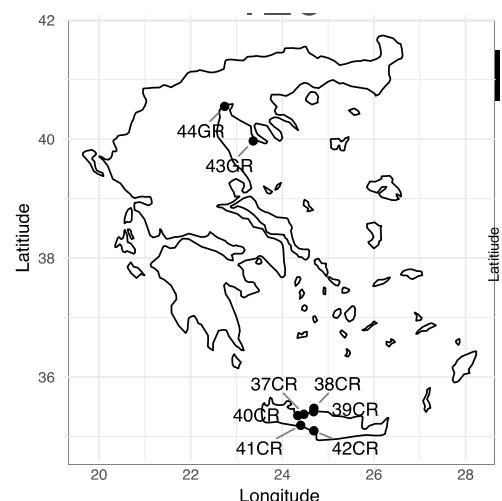
Likelihood($X|K$)



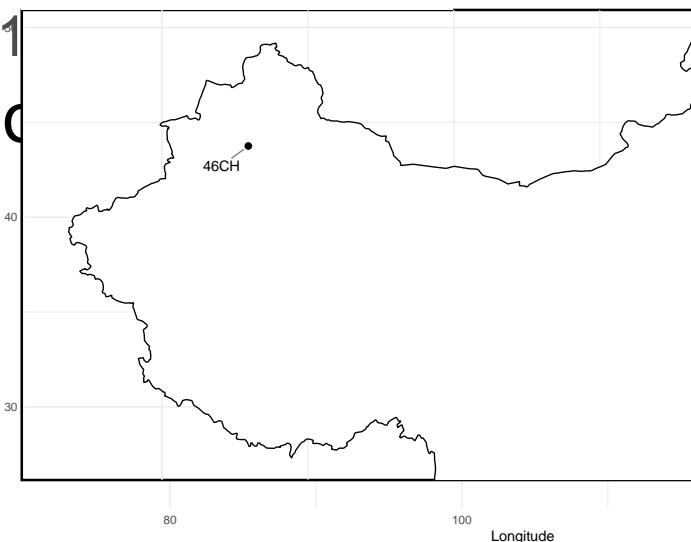
Southwest United States



Mainland
Greece & Crete



Xinjiang, China



Collected field samples from lab cultures and 36 sites within introduced and native range

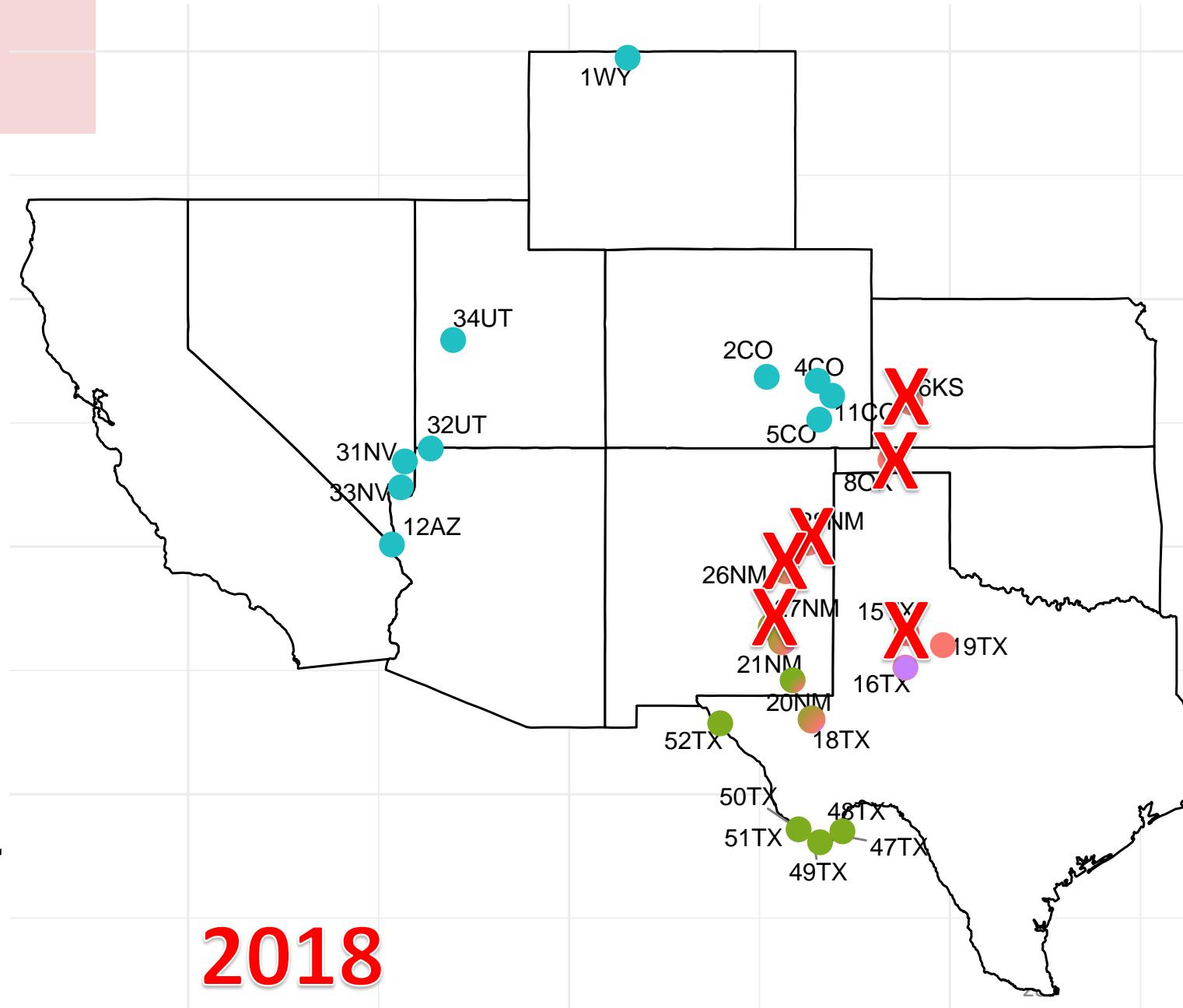


Prospects

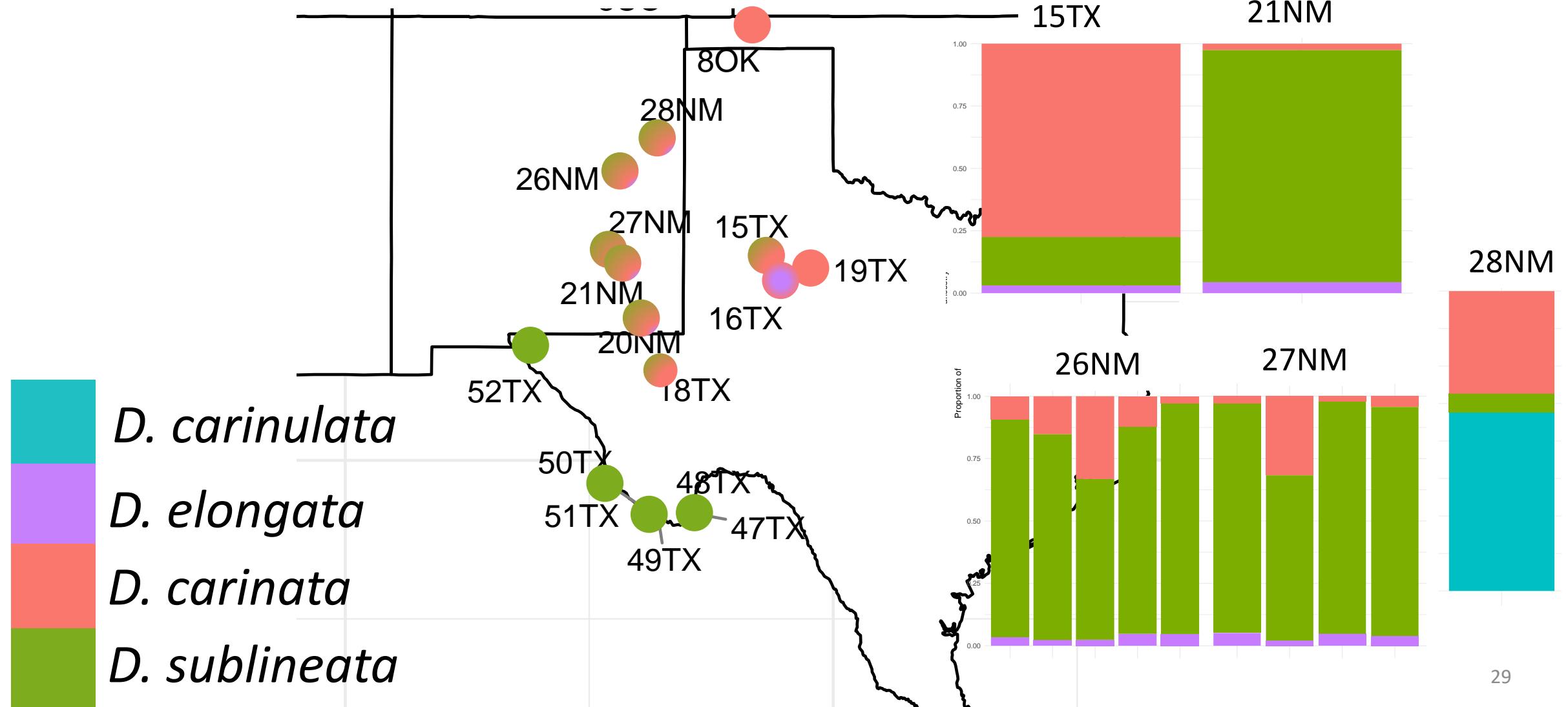
Hybrids and "pure" populations were abundant in 2014.

Neither hybridization status nor genomic diversity alone seem to predict establishment and spread.

2018



Ancestry assignment reveals tri-specific hybrids



Biocontrol Agents

Ibiocontrol.com 8/21/18

Subject Name	Scientific Name
flea beetle	<i>Altica carduorum</i>
brown dot leafy spurge flea beetle	<i>Aphthona cyparissiae</i>
black leafy spurge flea beetle	<i>Aphthona czwalinae</i>
copper leafy spurge flea beetle	<i>Aphthona flava</i>
brown-legged spurge flea beetle	<i>Aphthona lacertosa</i>
black dot spurge flea beetle	<i>Aphthona nigricutis</i>
flea beetles	<i>Aphthona</i> spp.
broom seed beetle	<i>Bruchidius villosus</i>
thistle tortoise beetle	<i>Cassida rubiginosa</i>
lesser St. John's wort beetle	<i>Chrysomela hyperici</i>
greater St. John's wort beetle	<i>Chrysomela quadrigemina</i>
leaf beetle	<i>Chrysomela</i> spp.
Mediterranean tamarisk beetle	<i>Diorhabda elongata</i>
black-margined loosestrife beetle	<i>Galerucella calmariensis</i>
golden loosestrife beetle	<i>Galerucella pusilla</i>
leaf beetles	<i>Galerucella</i> spp.
ragwort flea beetle	<i>Longitarsus jacobaea</i>

A reference genome allows us to anchor short reads, provides gene-specific context, and serves as a resource to other species

Predicted ranges (Tracy & Robbins 2009)

Diorhabda spp. Status

- *D. carinulata*, established
- ◊ *D. carinulata*, unestablished
- *D. elongata*, established
- ☆ *D. elongata*, little established
- ✚ *D. elongata*, caged/released
- ▲ *D. elongata*, unestablished
- *D. carinata*, caged/released
- ◇ *D. sublineata*, released

Habitat Suitability Index Models

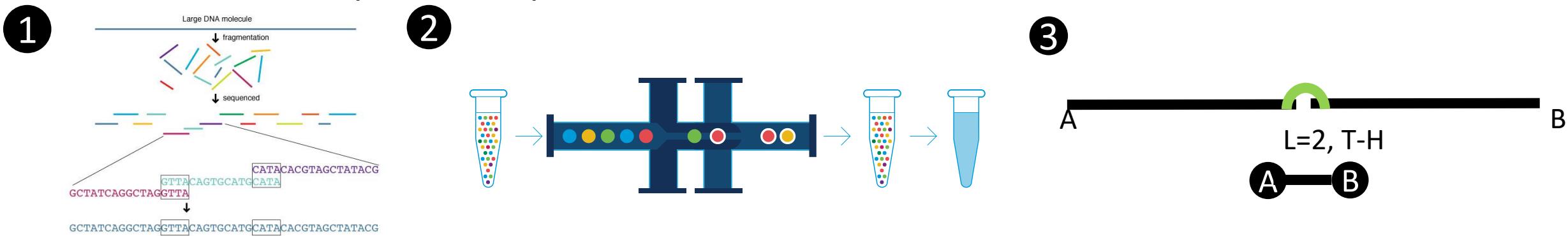
Scoring among Top 15%

- *D. elongata*
- *D. carinata*
- *D. sublineata*
- *D. carinulata*

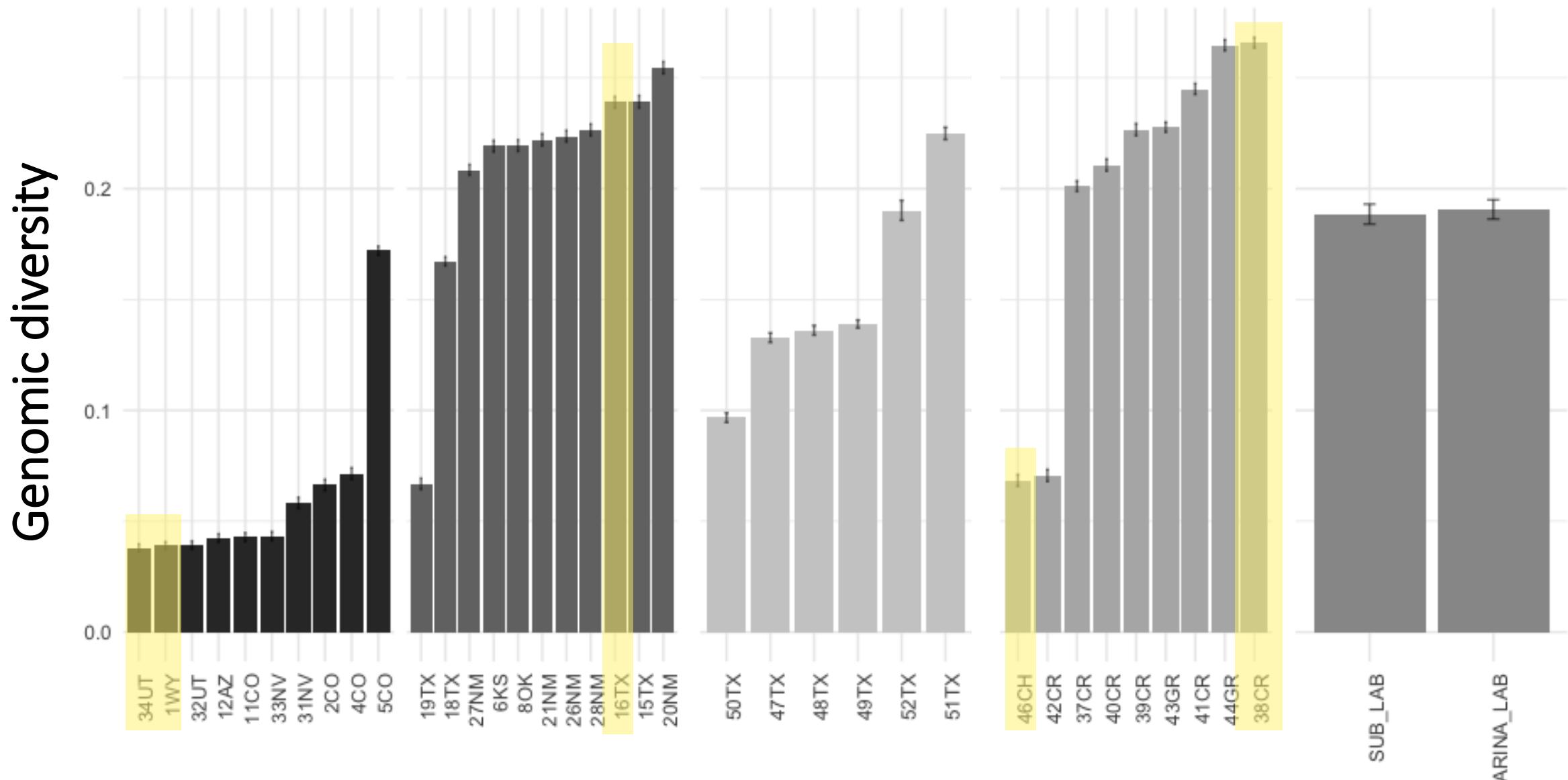


Cost-effective assembly of (one of) the first weed biocontrol (insect) agent reference genome (coming soon!)

Input data	③	① Shotgun	② 10X
ng DNA		2400	200
Cost		\$2,700	\$3,300
# contigs		21,901	79
Largest contig (Mbp)		.575	1.87
Total length (Mbp)		368.69	20.90
N50 (Kbp)		5.57	1.05
# N's per 100 kbp		40.11	2707.62

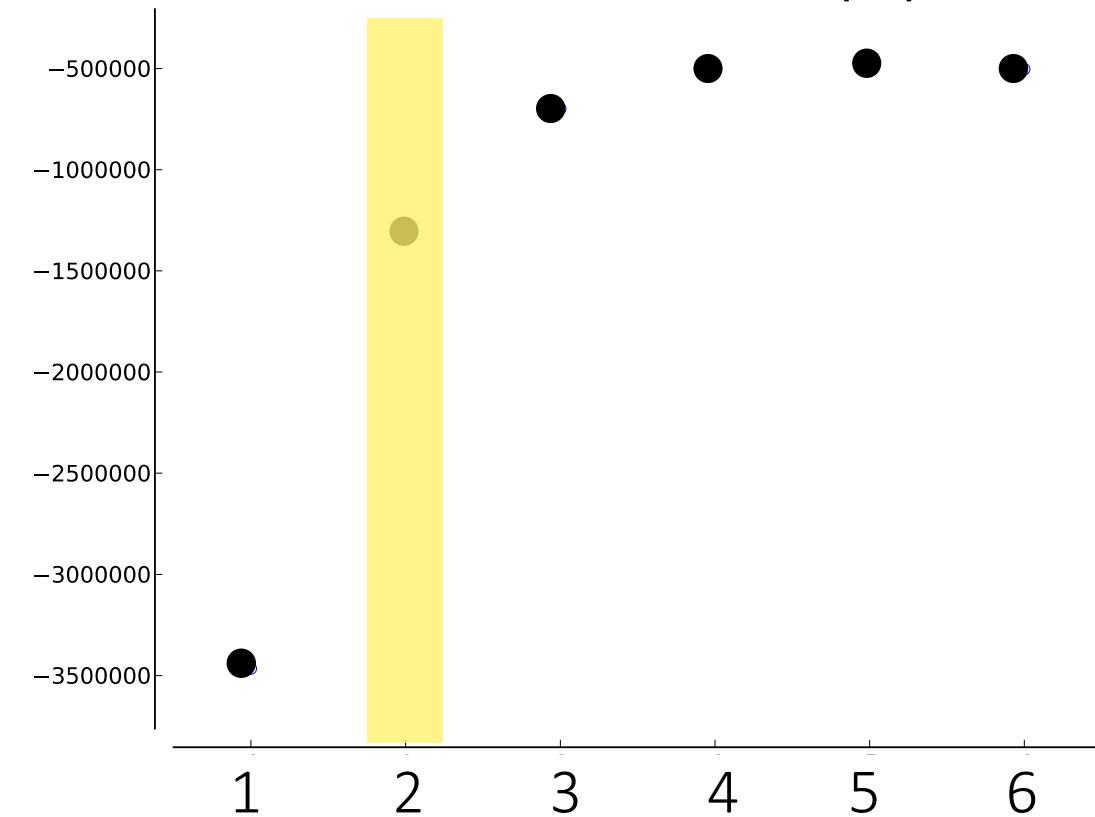


Introduced
D. carinulata Hybrid range
 hybrids No hybrids Native range Lab



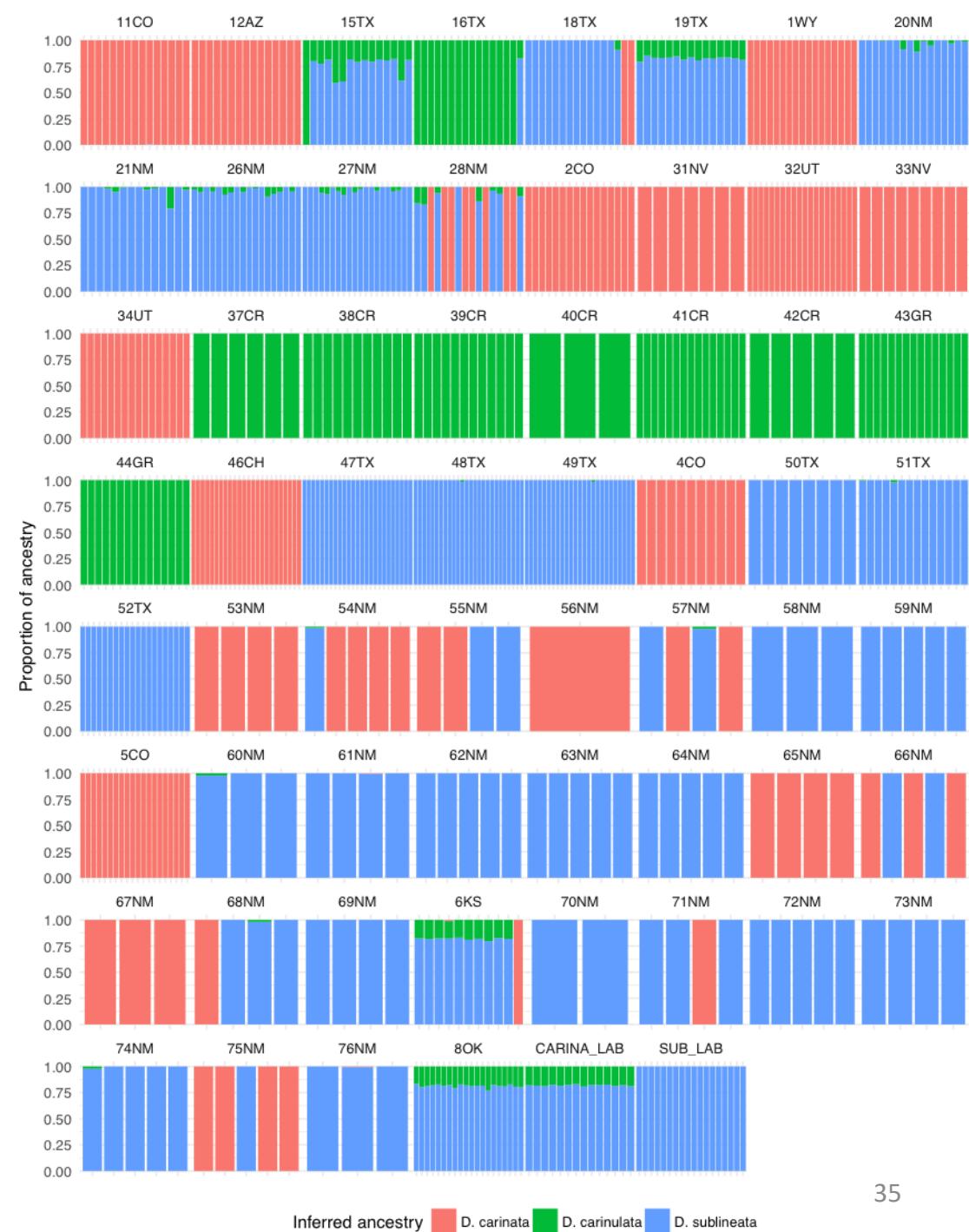
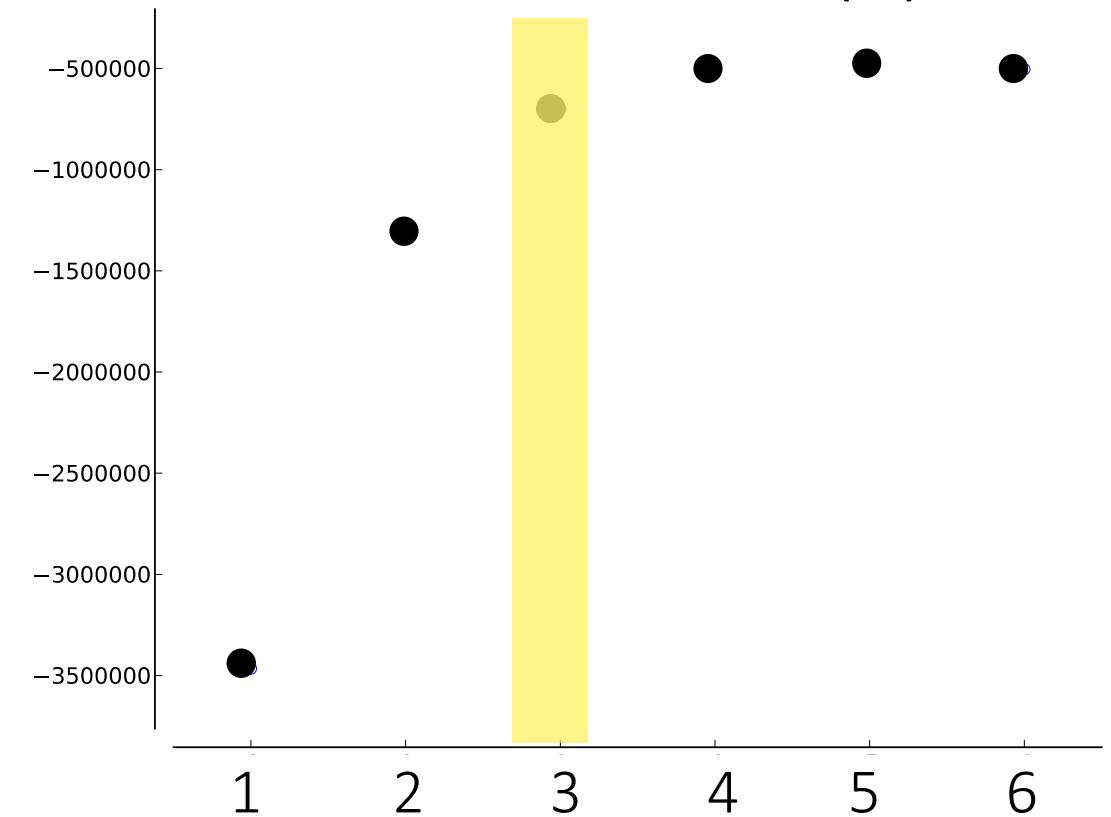
Hierarchical population structure reflects inter-specific relationships

Likelihood(K)

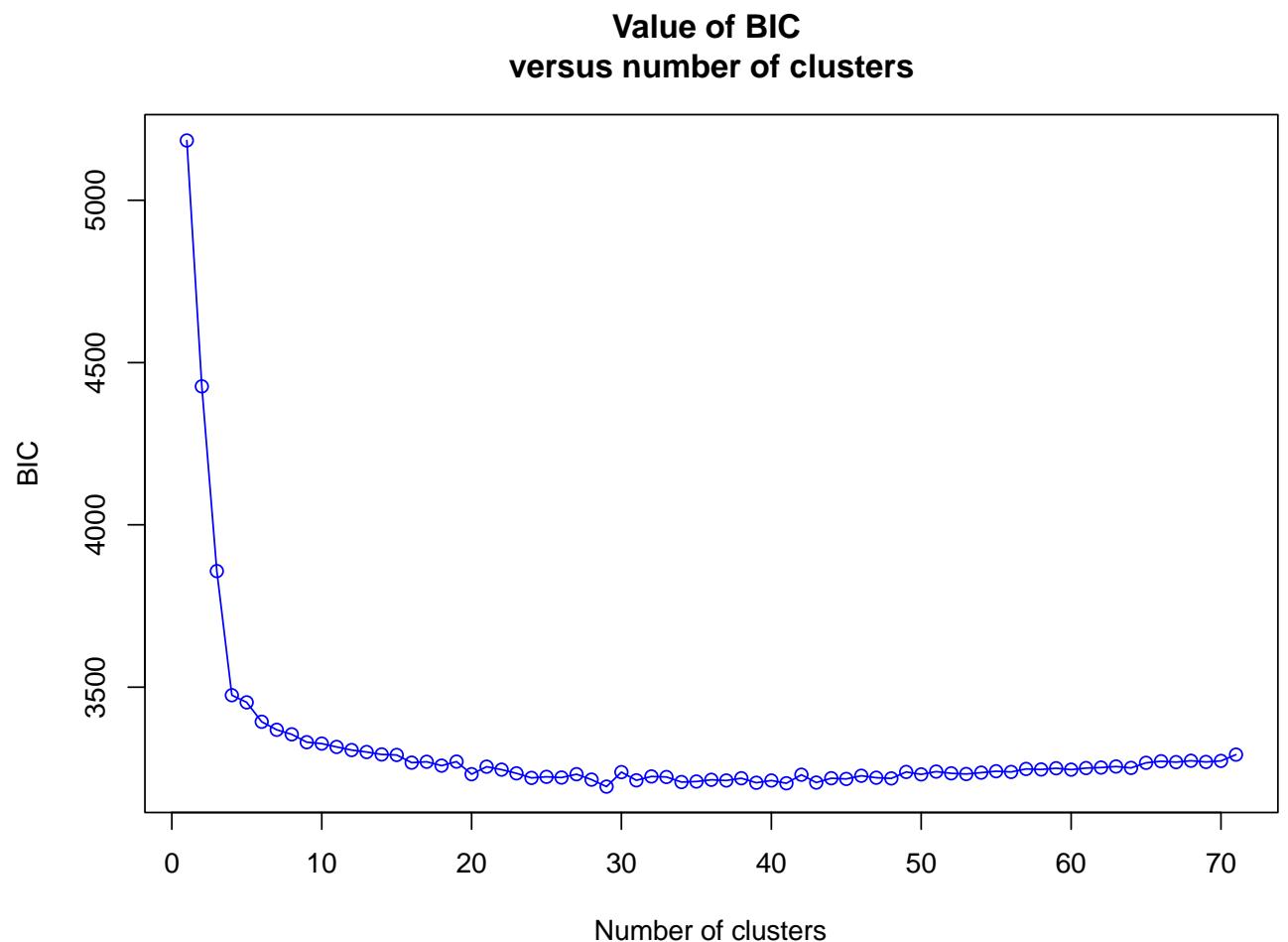
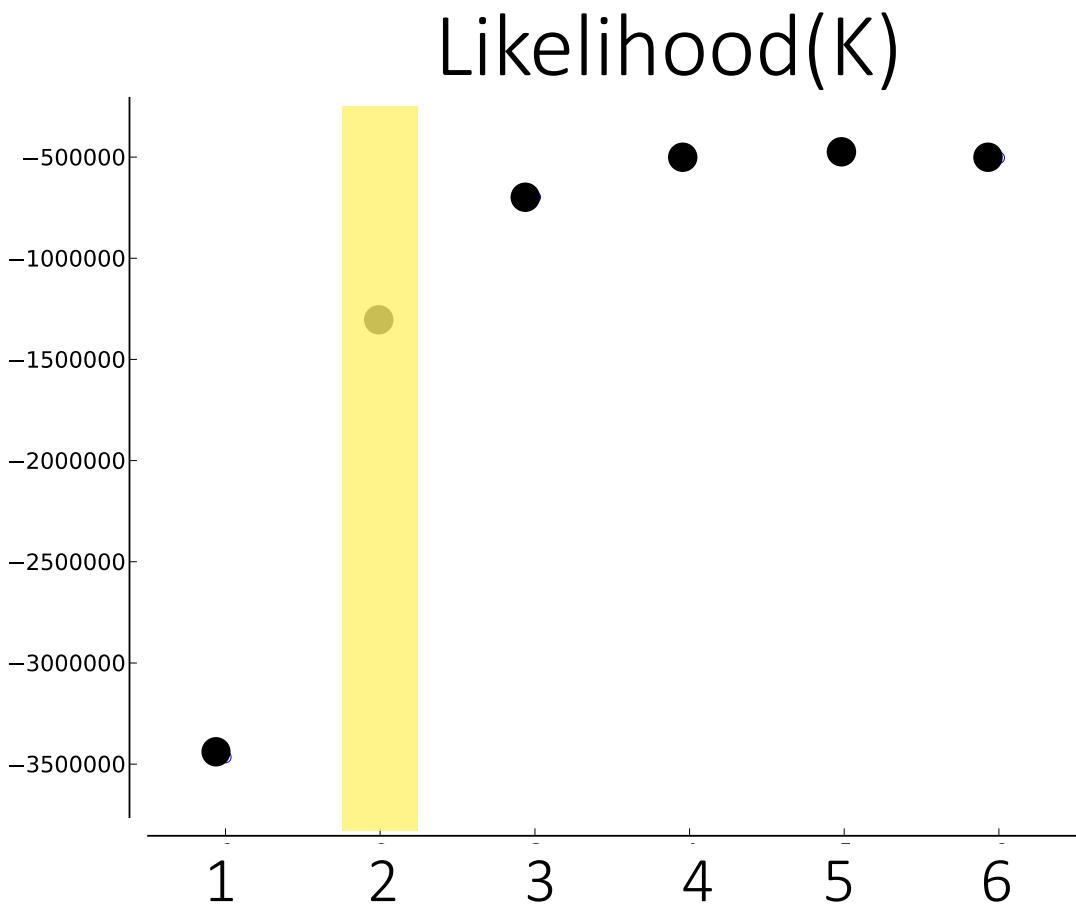


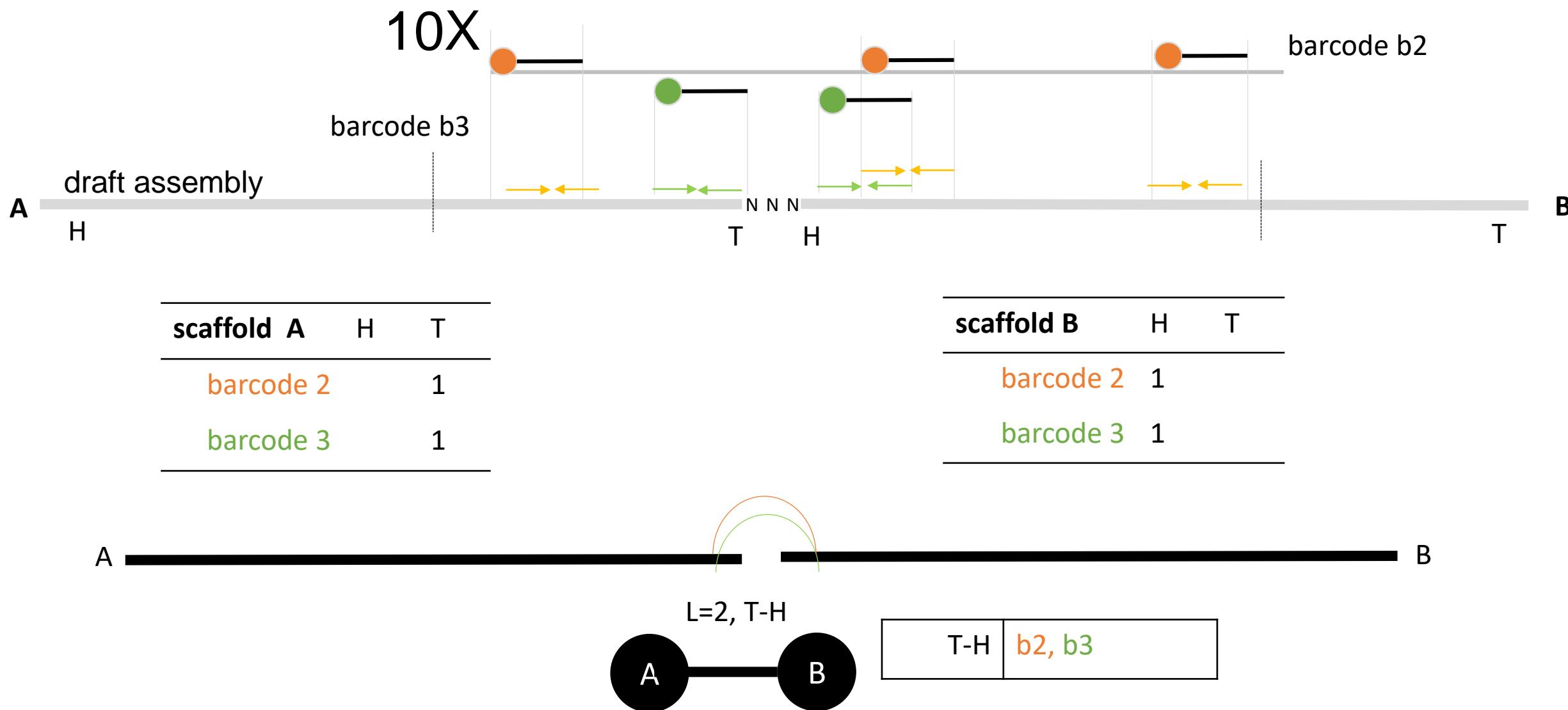
Hierarchical population structure reflects inter-specific relationships

Likelihood(K)



Choosing K





RADseq capitalizes on existing restriction enzyme technology, PCR, and high-throughput sequencing

